

Original Article

Plasma Glycosaminoglycans: A new Promising tool for Assessment of Non-Metastatic Renal Cell Carcinoma Patients Following Nephrectomy

Awais Ali^{1*}, Abdelkarem Omneya², Kashif Adil³

¹Department of Biochemistry, Abdul Wali Khan University Mardan, Mardan, Pakistan

²Department of Chemical Pathology, Medical Research Institute, Alexandria University, Alexandria, Egypt.

³Department of Urology, Hayatabad Medical Complex Hospital, Pakistan

Received: 18 April, 2024; Accepted: 16 June, 2024

DOI: 10.22037/nbm.v12i4.45088

Abstract

Background: Non-invasive detection of renal cell carcinoma (RCC) recurrence is a major challenge that could radically affect patient survival. To date, there are no approved biomarkers for inclusion in the monitoring and follow-up of RCC; therefore, assessment of treatment response is lacking.

Materials and Methods: A Cross-sectional study was carried out on biopsy-proven renal cell carcinoma patients scheduled for nephrectomy at Hayatabad Medical Complex (HMC) Hospital, Peshawar, Pakistan, between September 2022 and January 2024.

Results: A total of 160 samples were included in the study. Eighty samples were obtained from biopsy-proven non-metastatic renal cell carcinoma patients, of which 40 were collected pre-nephrectomy, 40 were collected post-nephrectomy from the same patients, and 80 samples were collected from age and gender-matched healthy individuals. Total plasma glycosaminoglycans (GAGs) levels were analyzed through a manual enzyme-linked immunosorbent assay using a non-competitive sandwich technique. Quantitative variables were summarized as means and standard deviation, while qualitative variables were summarized as frequency and percentage. A paired t-test was performed to check whether the difference between the mean plasma GAG levels in pre-nephrectomy and post-nephrectomy groups was significant.

Conclusion: The results revealed that post-nephrectomy GAG levels have significantly decreased compared to pre-nephrectomy levels ($P < 0.001$). Plasma glycosaminoglycan levels could be promising markers for monitoring renal cell carcinoma patients post-nephrectomy.

Keywords: Glycosaminoglycans, Nephrectomy, Renal cell carcinoma

*Corresponding Author: Awais Ali, Department of Biochemistry, Abdul Wali Khan University Mardan, Mardan, Pakistan. Email: awaisali@awkum.edu.pk. ORCID: <https://orcid.org/0000-0002-4514-9509>

Please cite this article as: Ali A, Omneya A, Adil K. Plasma Glycosaminoglycans: A new promising tool for assessment of non-metastatic Renal cell carcinoma patients following nephrectomy. Novel Biomed. 2024;12(4):142-8.

Introduction

Renal cell carcinoma (RCC) stands as the most common form of kidney malignancy, contributing to

approximately 179,000 deaths each year globally. According to the data of GLOBOCAN in 2020, kidney cancer is the 14th most common malignancy worldwide, with an estimated about 431 thousand patients newly

diagnosed in 2020¹. Kidney Cancer (KC) ranks as the sixth most prevalent cancer in males and the tenth in females on a global scale². Pakistan ranks eighth among the countries with a high rate of kidney diseases, with 17 million people suffering from kidney problems³. However, there are no updated statistics about the incidence of kidney cancer cases.

Several factors contribute to increased risk for RCC, including male sex, old age, central adiposity, tobacco smoking, alcohol intake, persistent hemodialysis, high blood pressure, and polycystic kidney disease⁴. Additionally, advances in the genetic landscape have recognized that both germline and somatic mutations could contribute to the development of KC. According to the Cancer Genome Atlas analysis, it's estimated that approximately 6–9% of submitted KC cases revealed a germline alteration in genes linked to cancer predisposition. However, due to the lack of extensive population-wide studies, researchers have overlooked much data regarding potentially influential autosomal recessive factors and gene variants that impact KC^{4–9}.

Around 75% of individuals diagnosed with renal cell carcinoma (RCC) have clear cell subtype (ccRCC), which commonly begins in the tubular epithelium of the kidney and has the potential to spread to various organs such as the liver, bones, lungs, and brain¹⁰.

Over 40% of RCC patients have no overt symptoms^{11–13}. RCC is mainly asymptomatic, with reports indicating that 20–40% of cases are diagnosed at the metastatic stage, which is universally deemed incurable^{14,15}.

The primary treatment approach for patients with non-metastatic RCC is surgical resection, either through partial or radical nephrectomy (M0 RCC). Nevertheless, around one-third of all M0 RCC patients encounter recurrence within five years following surgery¹¹. Despite the implementation of contemporary targeted therapies, patients with metastatic RCC still experience notably worse median survival rates¹⁶. Existing follow-up protocols focus on early detection of recurrence because symptomatic recurrences typically carry a worse prognosis than those detected through routine follow-up^{17,18}. Current follow-up protocols primarily depend on radiological imaging, typically involving chest and abdomen computed tomography every 6–12 months for 3–5

years, depending on the recurrence risk post-surgery¹⁹. Non-invasive detection of recurrence is a major unmet clinical need that could improve follow-up. To date, the absence of approved biomarkers in clinical management leads to delayed diagnosis or insufficient evaluation of treatment response. Although recent technologies involving methylated cfDNA and metabolomics show promise as markers, these assays are complex, costly, and lack standardized kits^{20,21}. Therefore, there is a need for an early non-invasive biomarker for diagnosis, disease surveillance, and RCC monitoring after therapy.

The extracellular matrix (ECM) constitutes a significant portion of the acellular compartment within the tumor microenvironment (TME), comprising major components such as proteoglycans (PGs) and hyaluronan (HA). A substantial portion of PGs is composed of repetitive chains known as glycosaminoglycans²².

Interestingly, plasma GAGs are emerging as a promising marker for diagnosing and monitoring surgically treated RCC patients. Glycosaminoglycans are structurally diverse heteropolysaccharides with viscous, lubricating characteristics similar to mucous secretions; therefore, they are also known as mucopolysaccharides¹⁹. These molecules can be located within the extracellular matrix or on the surfaces of animal cells. They have been discovered to bind and regulate various proteins, such as cytokines, chemokines, enzymes, growth factors (GF), morphogens, and adhesion molecules²⁰. The complex array of GAG sulfation and epimerization patterns, referred to as the GAGome, is increasingly recognized as a non-genetic code governing numerous biological functions, many of which play a role in cancer. Additionally, GAGs can bind to various ligands and are essential mediators of homeostasis and other pathological processes like tumorigenesis, wound healing defects, and blood clotting disorders^{21,22}. In Renal cell carcinoma patients, GAG excretion in the urine is considered a reliable diagnostic marker for determining tumor size and unilocularity or multilocularity²³.

Based on the importance of GAG levels in diagnosing RCC, our study aimed to evaluate plasma GAG levels as non-invasive biomarkers for monitoring post-nephrectomy RCC patients.

Methods

Study design: A cross-sectional study was conducted at Hayatabad Medical Complex (HMC) Hospital, Peshawar, Pakistan, between September 2022 and January 2024, to assess the role of glycosaminoglycans in monitoring patients with RCC post-nephrectomy (Ethical code: HMC/PMC-EB/MS/00343). The study was carried out according to the Helsinki Declaration 2013 and was approved by the HMC ethical committee. Patients diagnosed with biopsy-proven non-metastatic renal cell carcinoma scheduled for nephrectomy were enrolled in the study. A total of 160 samples were included in the study. Eighty samples were obtained from biopsy-proven renal cell carcinoma patients, of which 40 samples were collected pre-nephrectomy and 40 samples were collected post-nephrectomy from the same patients. Another 80 samples were collected from age and gender-matched healthy individuals as control samples.

Meanwhile, patients with urological cancer, bladder cell carcinoma, or any concurrent cancer were excluded. Written informed consents were obtained from all patients before participation in the study. All of the enrolled patients agreed to have their samples for research purposes.

GAG measurements: Three mL blood samples were collected from the study subjects in the antecubital vein and placed in an ethylenediaminetetraacetic acid (EDTA) vacutainer tube. The sample was then

processed by centrifugation at 2000-3000 RPM for approximately 20 minutes within two hours of sample collection, and the supernatant was collected and stored at -80 C until the analysis. Human total plasma GAG levels were analyzed through manual enzyme-linked immunosorbent assay (ELISA). The principle was based on a non-competitive sandwich ELISA technique, antigen-antibody interaction²⁴. The human GAG antibody was pre-coated on the plate. A GAG antigen in the sample was added, bound to the antibody, and coated on the well. The results of the samples were analyzed using an ELISA plate reader

Statistical analysis: Data were entered in a statistical software package (SPSS version 21). The normality of the data was tested using the Kolmogorov-Smirnov test. Mean and standard deviation were used to describe normally distributed quantitative variables. Qualitative variables were described as frequency and percentage. The paired t-test was used to compare normally distributed variables within dependent groups. A p-value of ≤ 0.05 was considered statistically significant.

Results

Patient characteristics: A total of 160 samples were included in the study. Eighty samples were obtained from biopsy-proven renal cell carcinoma patients, of which 40 samples were collected pre-nephrectomy and 40 samples were collected post-nephrectomy from the same patients. Another 80 samples were collected from age and gender-matched healthy individuals as control

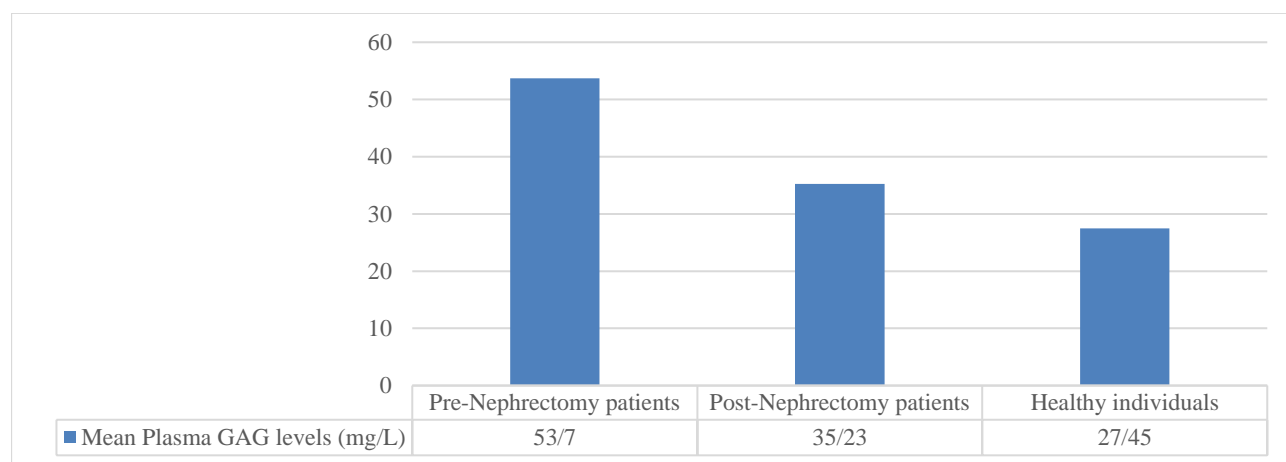


Figure 1. Mean plasma glycosaminoglycans levels in the studied groups.

samples.

The mean age of pre-operative patients is 59.50 ± 5.5 years compared to 58.60 ± 5.7 years for controls. The mean age of males was 59.24 ± 6.5 years, and the mean age of females was 60.24 ± 6.7 years. 26 (65 %) of the pre-operative patients were males, while 14 (35 %) were female. According to TNM staging, 39.6% of our RCC patients were T2NM0 stage while 24.3% were in the T3NM0 stage, and 36.1% were in the T4NM0 stage.

Plasma GAG levels in renal carcinoma patients pre and post-nephrectomy: The pre-nephrectomy mean plasma GAG levels was 53.70 ± 6.72 (mg/L) which was significantly higher than the post-nephrectomy mean plasma GAG levels 35.23 ± 5.28 (mg/L), ($t=19.31$, $P<0.001$). The mean plasma GAG levels of the healthy individual were 27.45 ± 4.82 (mg/L). Plasma GAG levels were significantly higher in pre-operative and postoperative samples than controls ($p<0.001$ and $p=0.03$). There was no significant difference in plasma GAG levels between males and females, whether pre-operative or postoperative ($p>0.05$).

Discussion

The discovery of promising biomarkers for monitoring cancer prognosis is one of the most significant challenges in oncologic research. Recently, numerous biomarkers have been demonstrated to reliably predict poor cancer prognosis through alterations in the concentration of proteins or small molecules²⁵⁻²⁷.

Our study aimed to assess the role of plasma GAGs as a non-invasive biomarker for monitoring RCC patients' response post-nephrectomy. Our study included 160 samples, 80 from histologically proven renal cell carcinoma patients, of which 40 samples were obtained pre-nephrectomy and 40 were obtained post-nephrectomy from the same patients. Another 80 samples were collected from age and gender-matched healthy individuals as control samples. According to TNM staging, 39.6% of our RCC patients were T2NM0 stage, while 24.3% were in the T3NM0 stage, and 36.1% were in the T4NM0 stage.

Our results showed that the mean of pre-operative plasma GAG levels was 53.70 ± 6.72 (mg/L), which

was significantly higher than the postoperative plasma GAG levels 35.23 ± 5.28 (mg/L), ($t=19.31$, $P<0.001$). The mean plasma GAG levels were significantly higher in pre-nephrectomy patients compared to healthy individuals (controls), confirming the role of plasma GAG levels as diagnostic marker for RCC. Additionally, the significant reduction in plasma GAG levels in post-nephrectomy patients could be explained as an early reaction towards the tumor. This underscores the significance of plasma GAG levels as a non-invasive marker for monitoring RCC patients post-nephrectomy. Our findings were consistent with Gatto et al. (2018) who reported that GAG scores were elevated in surgically treated RCC patients compared to healthy individuals, irrespective of the stage, grade, histology, age, or gender²⁸. Additionally, they concluded that GAGs are susceptible diagnostic and promising markers indicators in surgically managed RCC, regardless of stage, grade, or histological characteristics.

Interestingly, another study conducted by Gatto et al. (2023), showed a correlation between the GAG scores and progression-free survival (PFS) and overall survival (OS) in a prospective cohort of clear cell renal cell carcinoma (ccRCC) patients previously enrolled in another study^{29,30}.

Remarkably, Gatto et al. (2022) observed that plasma and urine-free GAGs are correlated with postsurgical recurrence in M0 RCC, as reported in a single-center prospective cohort study²⁹.

Additionally, changes in GAG profile were observed in metastatic clear cell (mcc)RCC patients. Moreover, they developed three GAG scores that facilitated precise identification of mccRCC patients. They successfully confirmed the score's reliability using an independent group of 18 patients with mccRCC and nine healthy individuals³¹.

Our results showed that plasma GAG levels were significantly higher in a post-nephrectomy group than controls ($p=0.03$); the timing of the post-nephrectomy sampling could explain this within 24 hours after the surgical procedure. However, Gatto et al., reported that glycosaminoglycans scores normalized in eight disease-free patients; however, these cases were assessed many years after the surgery. Therefore, frequent measurement of post-nephrectomy plasma GAGs is recommended to detect the accurate time of

normalization of plasma GAGs³¹.

Importantly, It was found that GAGs are completely changed in RCC tissue compared to normal adjacent tissues, confirming the tumor's effect in modifying GAG profile and supporting the fact that serum GAG levels play a crucial role in RCC diagnosis²⁶.

Currently, there is no approved biomarker for metastatic clear cell RCC in routine practice. Changes in the GAG profile are considered promising biomarkers for monitoring RCC patients and early detection of recurrence. Likewise, our findings suggest that plasma GAG levels could have clinical relevance in the monitoring and follow-up of non-metastatic RCC.

These biomarkers offer numerous potential benefits. Firstly, plasma GAGs offer diagnostic advantages for patients with non-metastatic RCC, as non-invasive biomarkers allow for dynamic disease monitoring. Secondly, the biological significance of GAGs supports their use as early biomarkers of cancer recurrence being involved in extracellular matrix interactions with chemokines activation. Both processes are crucial for tumor metastasis²³.

Despite the strong potential of plasma GAG levels as promising biomarkers for monitoring RCC patients, our study has some limitations. Firstly, we did not investigate the urine GAG levels post-nephrectomy in the enrolled patients. Secondly, our study is a Cross-sectional study with a relatively small sample size, therefore conducting further studies in prospective cohort design with larger sample size is highly encouraged. Finally, further prognostic studies, such as survival analysis, are needed to confirm the role of plasma GAG levels as a prognostic biomarker to guide patient risk stratification and clinical decision-making.

Some challenges in using GAGs as biomarkers need to be addressed in future studies. Firstly, there is a need for commercially standardized assays. Secondly, the a need for investigating the effect of ethnic variation and lifestyle factors on plasma GAG baseline levels in different populations.

Conclusion

Total Plasma GAGs are promising biomarkers for monitoring non-metastatic RCC patients.

Post-nephrectomy. The non-invasive nature of plasma GAGs and simple method of detection will facilitate frequent monitoring of such patients and early detection of cancer recurrence with subsequent improvement in the patient-centered care. Future studies to validate our results in multi-centric studies in different ethnic groups are highly recommended.

Authors' contributions

Awais Ali contributed to the research idea data collection, performed the laboratory work, interpreted the results, reviewed the literature, and wrote the manuscript.

Abdelkarem Omneya contributed to the research design, reviewed the laboratory work and results interpretation, reviewed the literature, and wrote the manuscript.

Kashif Adil contributed to the study participants' research design, recruitment, and clinical assessment and wrote the manuscript.

All authors have read, edited, and approved the final version of the manuscript.

Acknowledgment

None.

Funding

None.

Conflict of interest

The authors declare that they have no conflict of interest.

References

1. Sung H, Ferlay J, Siegel RL, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin.* 2021;71:209–49.
2. Peired AJ, Campi R, Angelotti ML, et al. Sex and gender differences in kidney cancer: clinical and experimental evidence. *Cancers (Basel).* 2021;13:4588.

3. Zaman S. 17 million Pakistanis suffering from kidney diseases.
4. Manzoor U, Ali A, Ali SL, et al. Mutational screening of GDAP1 in dysphonia associated with Charcot-Marie-Tooth disease: clinical insights and phenotypic effects. *J Genet Eng Biotechnol.* 2023;21:1–11.
5. Huang K, Mashl RJ, Wu Y, et al. Pathogenic germline variants in 10,389 adult cancers. *Cell.* 2018;173:355–70.
6. Miriam N, Olayinka F, Olufunbi A, et al. Effect of Ethanolic *Ocimum Tenuiflorum* (Holy Basil) Extract on Diclofenac Induced Hepatotoxicity in Rats.
7. ALi A, Manzoor U, Ali SL, et al. Currently Trending and Futuristic Biological Modalities in The Management of Different Types of Diabetes: A Comprehensive Review. *J Popul Ther Clin Pharmacol.* 2023;30:2948–70.
8. Ali SL, Ali A, Alamri A, et al. Genomic annotation for vaccine target identification and immunoinformatics-guided multi-epitope-based vaccine design against Songling virus through screening its whole genome encoded proteins. *Front Immunol.* 2023;14:1284366.
9. Nwanna E, Ojo R, Shafiq N, et al. An In Silico In Vitro and In Vivo Study on the Influence of an Eggplant Fruit (*Solanum anguivi* Lam) Diet on Metabolic Dysfunction in the Sucrose-Induced Diabetic-like Fruit Fly (*Drosophila melanogaster*). *Foods.* 2024;13:559.
10. Kase AM, George DJ, Ramalingam S. Clear cell renal cell carcinoma: from biology to treatment. *Cancers (Basel).* 2023;15:665.
11. Sun M, Choueiri TK. Recurrence in renal cell carcinoma: the work is not done. *Nat Rev Urol.* 2016;13:246–7.
12. Saleem Naz Babari I, Islam M, Saeed H, et al. Design, synthesis, in-vitro biological profiling and molecular docking of some novel oxazolones and imidazolones exhibiting good inhibitory potential against acetylcholine esterase. *J Biomol Struct Dyn.* 2024;1–18.
13. Ali SL, Ali A, Alamri A. Genomic annotation for vaccine target identification and multi-epitope-based vaccine design against Songling virus through screening its whole genome encoded proteins. DOI: 10.3389/fimmu. 2023.1284366.
14. Belldegrun AS, Klatter T, Shuch B, et al. Cancer-specific survival outcomes among patients treated during the cytokine era of kidney cancer (1989-2005) A benchmark for emerging targeted cancer therapies. *Cancer.* 2008;113:2457–63.
15. Dabestani S, Thorstenson A, Lindblad P, et al. Renal cell carcinoma recurrences and metastases in primary non-metastatic patients: a population-based study. *World J Urol.* 2016;34:1081–6.
16. Gupta K, Miller JD, Li JZ, et al. Epidemiologic and socioeconomic burden of metastatic renal cell carcinoma (mRCC): a literature review. *Cancer Treat Rev.* 2008;34:193–205.
17. Dabestani S, Beisland C, Stewart GD, et al. Long-term outcomes of follow-up for initially localised clear cell renal cell carcinoma: RECUR database analysis. *Eur Urol Focus.* 2019;5:857–66.
18. Shafiq N, Arshad M, Ali A, et al. Integrated computational modeling and in-silico validation of flavonoids-Alliucide G and Alliucide A as therapeutic agents for their multi-target potential: Combination of molecular docking, MM-GBSA, ADMET and DFT analysis. *South African J Bot.* 2024;169:276–300.
19. Beisland C, Guðbrandsdóttir G, Reisæter LAR, et al. A prospective risk-stratified follow-up programme for radically treated renal cell carcinoma patients: evaluation after eight years of clinical use. *World J Urol.* 2016;34:1087–99.
20. Nuzzo PV, Berchuck JE, Korthauer K, et al. Detection of renal cell carcinoma using plasma and urine cell-free DNA methylomes. *Nat Med.* 2020;26:1041–3.
21. Li H, Bullock K, Gurjao C, et al. Metabolomic adaptations and correlates of survival to immune checkpoint blockade. *Nat Commun.* 2019;10:1–6.
22. Wieboldt R, Läubli H. Glycosaminoglycans in cancer therapy. *Am J Physiol Physiol.* 2022;322:1187–200.
23. Afratis N, Gialeli C, Nikitovic D, et al. Glycosaminoglycans: key players in cancer cell biology and treatment. *FEBS J.* 2012;279:1177–97.
24. Dinh P, Tran C, Dinh T, et al. Hsa_circRNA_0000284 acts as a ceRNA to participate in coronary heart disease progression by sponging miRNA-338-3p via regulating the expression of ETS1. *J Biomol Struct Dyn.* 2023;1–14.
25. Jonasch E, Futreal PA, Davis IJ, et al. State of the science: an update on renal cell carcinoma. *Mol cancer Res* 2012; 10: 859–880.
26. Moch H, Srigley J, Delahunt B, et al. Biomarkers in renal cancer. *Virchows Arch.* 2014;464:359–65.
27. Parker AS, Eckel-Passow JE, Serie D, et al. Higher expression of topoisomerase II alpha is an independent marker of increased risk of cancer-specific death in patients with clear cell renal cell carcinoma. *Eur Urol.* 2014;66:929–935.
28. Gatto F, Blum KA, Hosseini SS, et al. Plasma glycosaminoglycans as diagnostic and prognostic biomarkers in surgically treated renal cell carcinoma. *Eur Urol Oncol.* 2018;1:364–77.
29. Gatto F, Dabestani S, Bratulic S, et al. Plasma and urine free glycosaminoglycans as monitoring biomarkers in non-metastatic renal cell carcinoma—a prospective cohort study. *Eur Urol Open Sci.* 2022;42:30–9.
30. Gatto F, Bratulic S, Jonasch E, et al. Plasma and Urine Free Glycosaminoglycans as Monitoring and Predictive Biomarkers in Metastatic Renal Cell Carcinoma: A Prospective Cohort Study. *JCO Precis Oncol.* 2023;7:e2200361.
31. Gatto F, Volpi N, Nilsson H, et al. Glycosaminoglycan profiling in patients' plasma and urine predicts the occurrence of metastatic clear cell renal cell carcinoma. *Cell Rep.* 2016;15:1822–36.
19. Yoo JY, Groer M, Dutra SV, Sarkar A, McSkimming DI. Correction: Yoo, JY, et al. Gut Microbiota and Immune System Interactions. *Microorganisms* 2020, 8, 1587. *Microorganisms.* 2020;8(12):2046.
20. Mazkour S, Shekarforoush SS, Basiri S. The effects of supplementation of *Bacillus subtilis* and *Bacillus coagulans* spores on the intestinal microflora and growth performance in rat. *Iranian journal of microbiology.* 2019;11(3):260.
21. Adibpour N, Hosseini-zhad M, Pahlevanlo A, Hussain MA. A review on *Bacillus coagulans* as a Spore-Forming Probiotic. *Applied Food Biotechnology.* 2019;6(2):91-100.
22. Azimirad M, Alebouyeh M, Naji T. Inhibition of lipopolysaccharide-induced interleukin 8 in human adenocarcinoma cell line HT-29 by spore probiotics: *B. coagulans* and *B. subtilis* (natto). *Probiotics and antimicrobial proteins.* 2017;9:56-63.

23. Noori M, Azimirad M, Eslami G, Looha MA, Yadegar A, Ghalavand Z, Zali MR. Surface layer protein A from hypervirulent *Clostridioides difficile* ribotypes induce significant changes in the gene expression of tight junctions and inflammatory response in human intestinal epithelial cells. *BMC microbiology*. 2022;22(1):259.
24. Lee JY, Lee JD, Phipps S, Noakes PG, Woodruff TM. Absence of toll-like receptor 4 (TLR4) extends survival in the hSOD1 G93A mouse model of amyotrophic lateral sclerosis. *Journal of neuroinflammation*. 2015;12:1-9.
25. Shinde T, Vemuri R, Shastri MD, Perera AP, Tristram S, Stanley R, Eri R. Probiotic *Bacillus coagulans* MTCC 5856 spores exhibit excellent in-vitro functional efficacy in simulated gastric survival, mucosal adhesion and immunomodulation. *Journal of Functional Foods*. 2019;52:100-8.
26. Clair G, Esbelin J, Mallea S, Bornard I, Carlin F. The spore coat is essential for *Bacillus subtilis* spore resistance to pulsed light, and pulsed light treatment eliminates some spore coat proteins. *International journal of food microbiology*. 2020;323:108592.
27. Freedman KE, Hill JL, Wei Y, Vazquez AR, Grubb DS, Trotter RE, Wrigley SD, Johnson SA, Foster MT, Weir TL. Examining the gastrointestinal and immunomodulatory effects of the novel probiotic *Bacillus subtilis* DE111. *International journal of molecular sciences*. 2021;22(5):2453.
28. Tobita K, Meguro R. *Bacillus subtilis* BN strain promotes Th1 response via Toll-like receptor 2 in polarized mouse M1 macrophage. *Journal of Food Biochemistry*. 2022;46(2):14046.
29. Permpoonpattana P, Hong HA, Khaneja R, Cutting SM. Evaluation of *Bacillus subtilis* strains as probiotics and their potential as a food ingredient. *Beneficial microbes*. 2012;3(2):127-36.
30. Tavares Batista M, Souza RD, Paccetz JD, Luiz WB, Ferreira EL, Cavalcante RC, Ferreira RC, Ferreira LC. Gut adhesive *Bacillus subtilis* spores as a platform for mucosal delivery of antigens. *Infection and immunity*. 2014;82(4):1414-23.
31. Hong HA, Duc LH, Cutting SM. The use of bacterial spore formers as probiotics. *FEMS microbiology reviews*. 2005;29(4):813-35.
32. Wang K, Yang X, Wu Z, Wang H, Li Q, Mei H, You R, Zhang Y. *Dendrobium officinale* polysaccharide protected CCl₄-induced liver fibrosis through intestinal homeostasis and the LPS-TLR4-NF- κ B signaling pathway. *Frontiers in Pharmacology*. 2020;11:240.
33. Fan J, Liu S, Ai Z, Chen Y, Wang Y, Li Y, et al. Fermented ginseng attenuates lipopolysaccharide-induced inflammatory responses by activating the TLR4/MAPK signaling pathway and remediating gut barrier. *Food & Function*. 2021;12(2):852-61.
34. Yu P, Ke C, Guo J, Zhang X, Li B. *Lactobacillus plantarum* L15 alleviates colitis by inhibiting LPS-mediated NF- κ B activation and ameliorates DSS-induced gut microbiota dysbiosis. *Frontiers in Immunology*. 2020; 11:575173.
35. Li T, Li F, Liu X, Liu J, Li D. Synergistic anti-inflammatory effects of quercetin and catechin via inhibiting activation of TLR4-MyD88-mediated NF- κ B and MAPK signaling pathways. *Phytotherapy Research*. 2019;33(3):756-67.
36. Wang Y, Lin J, Cheng Z, Wang T, Chen J, Long M. *Bacillus coagulans* TL3 Inhibits LPS-Induced Caecum Damage in Rat by Regulating the TLR4/MyD88/NF- κ B and Nrf2 Signal Pathways and Modulating Intestinal Microflora. *Oxidative Medicine and Cellular Longevity*. 2022;2022(1):5463290.