Original Article

Morphological Study of *Fasciola* Parasites Isolated from Cattle and Sheep in Golestan Province (Iran)

Ahmad Halakou¹, Hooshang Khazan²*, Mojgan Bendehpour³, Bahram Kazemi³

¹Ph.D Candidate of Parasitology Department, International Branch, Shahid Beheshti University of Medical Sciences, Tehran, Iran

²Department of Parasitology, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran.

³Department of Biotechnology, School of Advanced Technologies in Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran

Received: 31 December, 2016; Accepted: 04 March, 2017

Abstract

Background: The genus Fasciola parasite causes fascioliasis infection. Fascioliasis is widespread all around the world and it is finding in abundance in the northern provinces of Iran. Cattle and sheep are the main hosts of the Fasciola parasite and intermediate hosts are lymnaeid snails such as Galba and Fossaria. Two main species of this genus are F. *hepatica* and F. *gigantica*. One of the most important methods of diagnosing this worm is morphological method. The aim of this study is to identify Fasciola through the morphological method in Golestan province.

Materials and Methods: *Fasciola* worms taken from infected livestock livers were washed three times with PBS and were stained with carmine alum. After staining using Valero and Periago methods, the worms were measured morphologically by calibrated microscope, stereomicroscope, and True Chrome II camera. SPSS version 19 was used for analysis of the data.

Results: A total of 45 livers from infected sheep and cattle with *Fasciola* worms were taken out of 228 samples, including 84 *Fasciola hepatica* (36.18%), 117 *Fasciola gigantica* (51.31%) and 27 *Fasciola* sp. (11.84%).

Conclusion: This study showed that the two main species of worms that is *F.hepatica* and *F. gigantica* were found in abundance in Golestan province. The current study was unable to identify 11.84% genus *Fasciola* showed as *Fasciola sp*.

Keywords: Morphological, Fasciola, cattle, sheep and Golestan

*Corresponding Author: Hooshang Khazan, Department of Parasitology, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran. Tel: (+98) 21-23872564; Email: khazan_h36@yahoo.co.in

Please cite this article as: Halakou A, Khazan H, Bendehpour M, Kazemi B. Morphological Study of *Fasciola* Parasites Isolated from Cattle and Sheep in Golestan Province (Iran). Novel Biomed. 2017;5(4):166-171.

Introduction

Fascioliasis one of the important diseases in animals and humans caused by *genus Fasciola*, two pathogenic species of this genus are *F. hepatica* and *F. gigantica*¹. This illness has traditionally been considered as a veterinary problem that causes important economic losses thanks to its impact on livestock, especially sheep and $cattle^2$ and of only secondary impact on humans³.

Cattle and sheep are the main hosts of the *Fasciola* parasite and intermediate hosts are lymnaeid snails such as *Galba* and *Fossaria*⁴. This parasite is widespread around the world and in Iran *F. hepatica* and *F. gigantic* coexist^{5,6}. One of the most common methods for detection genus Fasciola in the 1970s was

based on morphological properties of the parasite such as fluke length, fluke breadth, cephalic cone length, cephalic cone breadth, testes length⁷.

In recent years, several studies were done by morphological method^{5,8,9,10}. In the past two decades, two large outbreaks of human fascioliasis occurred in Guilan province in northern Iran^{6,11} as well as one epidemic human fascioliasis was reported in the western province of Kermanshah in Iran^{12,13,14}. Several studies about fasciola parasite carried out in Guilan, Kermanshah and some other provinces in Iran^{12,15} but, few if any studies have been done in this field in Golestan province. To this end, the aim of this study was to do a morphological scrutiny and species identification of Fasciola parasites isolated from cattle and sheep in Golestan province.

Methods

Liver flukes were obtained from infected cattle and sheep in several slaughterhouses of Gorgan, Gonbad Kavus, Ramian, Azad shaher, Aliabad, Minoodasht, Aqqala, Bander Turkman and Kalaleh in Golestan province, in northeast of Iran. Figure 1 shows Golestan province and the locations where the samples were collected. Liver flukes collection was carried out daily in the region during a one year period (December 2014 to December 2015). The liver worms were carefully separated and fixed in formalin between a slide and cover glass then they were stained with carmine alum and mounted entellan¹⁶ and measured morphometrically by calibrated microscope, stereomicroscope, and True Chrome II camera, which was installed on them. All standardized measurements of flukes measured according to methods proposal by Valero and colleagues^{8,17,18}.

Recently, researchers have used the technique to identify morphometric fasciolids^{5,8,19}. Parameter measurements including: Body length (BL), Body width (BW), Cone length (CL), Cone width (CW), Oral sucker maximum diameter (OS_{max}), Oral sucker $(OS_{min}),$ minimum diameter Ventral sucker maximum diameter $(VS_{max}),$ Ventral sucker minimum diameter (VS_{min}), Distance between anterior end of body and VS (A-VS), Distance between suckers (OS-VS), Pharynx length (PhL), Pharynx width (PhW)^{5,8,19}.

SPSS (Statistical Package for Social Sciences) for windows version 19 was used for the analysis of the data. ANOVA test was used to determine the significance of difference among the groups, and t test was used to compare the two hosts in cattle and sheep (Table 1).

Results

Figures 2 and 3 demonstrate the measurements carried out with calibrated microscope (Leica Galen III) and stereomicroscope (Optica). The results of ANOVA showed that among factors measured there were significant differences among body length, body width, BL/BW ratio, Cone length, VS min and VS max while the following factors Cone width, A-VS, OS-VS, Ph L, Ph W, OS max and OS min were not significantly different (P≤0.05) (table 1). T test results showed a significant difference in the size of the body length, body width, BL/BW ratio, Cone length, Pharynx length, VS max and VS min worms isolated from cattle and sheep (P≤0.05).

In the current study, out of 31 infected livers of sheep and 14 infected livers of cattle, 228 flukes of *Fasciola* were collected altogether. With regard to variety of infection, five worms were collected from each infected liver on average. Based on morphological criteria in the method, 84 (36.84%) worms were identified as *Fasciola hepatica*, 117 (51.31%) as *Fasciola gigantica* and 27 (11.84) as *Fasciola sp*. The findings of this research were according to different towns and Fasciola species listed in Table 2. In all towns under investigation, two species of Fasciola were identified and isolated. In two locations that is, Aq Qala and Minoodasht *Fasciola gigantica* and *Fasciola hepatica* were not identified respectively.

In the study out of 31 infected livers of sheep 145 *Fasciola* worms were isolated from which 82 species were *F. hepatica* (56.55%), 39 species *F. gigantica* (26.89%) and 24 species were not identified correctly. Besides, out of 14 infected livers of cattle 83 *Fasciola* worms were isolated from which 78 species were *F. gigantica* (93.97%) and two species of *F. hepatica* (2.4%) and 3 species were not identified correctly (Table 3).

Table 1: Comparative morphological data of liver flukes for Sheeps and Cattle from Golestan Province in Iran. A-VS	:
Distance between anterior end of body and VS.	

Fasciola measurements (mm)	<i>₹. hepatica</i> in Sheep N=82	<i>F. hepatica</i> in Cattle N=2	<i>F. gigantica</i> in Sheep N=34	<i>F. gigantica</i> in Cattle N=83	Fasciola.sp in Sheep N=24	Fasciola.sp in Cattle N=3	P Value
Body length, BL	16.3 – 36.1 26.13 ±4.24	15.3 -19.9 17.6 ± 3.25	$\begin{array}{c} 16.2 - 39.2 \\ 30.82 \pm 5.06 \end{array}$	24.9 - 50.99 34.28 ± 7.19	16.1 - 34.2 25.41 ± 4.91	26.8 - 32.1 29.8 ± 2.73	.000
Body width, BW	7.8 – 17.4 12.26 ±2.42	7.1 - 8.1 7.6 ± 0.70	5.2 - 12.9 8.27 ± 1.93	4.22 - 10.1 7.03 ± 1.21	6.4 - 13.8 9.60 ± 2.08	5.9 - 8.1 7.06 ± 1.1	.000
BL/BW ratio	1.61 - 3.50 2.19 ± 0.44	1.8 - 2.8 2.3 ± 0.70	2.67 - 6.32 3.83 ± 0.84	2.64 - 9.76 5.08 ± 1.63	2.09 - 3.91 2.68 ± 0.44	3.96 - 4.54 4.25 ± 0.29	.000
Cone length, CL	$\begin{array}{r} 1.46 - 3.38 \\ 2.31 \pm \ 0.48 \end{array}$	$\begin{array}{c} 2.15 - 2.19 \\ 2.17 \pm 0.02 \end{array}$	$\begin{array}{c} 1.41 - 3.31 \\ 2.54 \pm 0.42 \end{array}$	1.65 - 4.45 2.9 ± 0.63	1.73 - 3.11 2.28 ± 0.41	2.55 - 3.28 2.85 ± 0.38	.000
Cone width, CW	$\begin{array}{c} 2.31 - 5.23 \\ 3.70 \pm 0.77 \end{array}$	2.99 - 3.02 3 ± 0.02	$\frac{1.92 - 4.62}{3.47 \pm 0.61}$	$\frac{1.4 - 4.41}{3.48 \pm 0.51}$	$\frac{1.90 - 4.39}{3.38 \pm 0.67}$	3.81 - 3.96 3.87 ± 0.07	.284
A-VS	$\begin{array}{c} 1.52 - 3.66 \\ 2.43 \pm 0.49 \end{array}$	$\begin{array}{c} 1.97 - 2.27 \\ 2.12 \pm 0.21 \end{array}$	1.58 - 3.21 2.21 ± 0.48	1.47 - 3.77 2.33 ± 0.46	1.07 - 3.19 2.21 ± 0.44	2.41 - 2.68 2.51 ± 0.14	.244
Distance between suckers (OS-VS)	0.99 - 2.88 1.79 ± 0.46	1.48 - 1.85 1.66 ± 0.26	0.6 - 2.44 1.53 ± 0.4	0.9 - 2.96 1.71 ± 0.43	0.73 - 2.55 1.59 ± 0.39	1.62 - 2.12 1.85 ± 0.25	.192
Pharynx length, PhL	0.31 - 1.03 0.67± 0.15	$\begin{array}{c} 0.35 - 0.37 \\ 0.36 \pm 0.01 \end{array}$	$\begin{array}{c} 0.37 - 0.98 \\ 0.64 \pm 0.15 \end{array}$	$\begin{array}{c} 0.32 - 1.15 \\ 0.6 \pm 0.13 \end{array}$	$\begin{array}{c} 0.49 - 0.99 \\ 0.63 \pm 0.17 \end{array}$	0.59 - 0.65 0.62 ± 0.03	.293
Pharynx width, PhW	0.27 - 0.88 0.44 ± 0.12	$\begin{array}{c} 0.22 - 0.31 \\ 0.26 \pm 0.06 \end{array}$	0.29 - 0.83 0.48 ± 0.12	$\begin{array}{c} 0.25 - 0.73 \\ 0.45 \pm 0.09 \end{array}$	0.21 - 0.77 0.41 ± 0.12	0.41 - 0.49 0.46 ± 0.04	.093
Oral sucker maximum diameter (OS max)	$\begin{array}{c} 0.51 - 1.22 \\ 0.82 \ \pm 0.16 \end{array}$	$\begin{array}{c} 0.78 - 0.84 \\ 0.81 \pm 0.04 \end{array}$	0.66 - 1.16 0.87 ± 0.13	0.55 - 1.06 0.83 ± 0.12	0.44 - 1.2 0.83 ± 0.2	$\begin{array}{c} 0.84 - 0.99 \\ 0.9 \pm 0.07 \end{array}$.416
Oral sucker minimum diameter (OS min)	$\begin{array}{c} 0.34 - 0.99 \\ 0.62 \pm 0.12 \end{array}$	0.38 - 0.64 0.51 ± 0.18	0.49 - 0.96 0.67± 0.13	$\begin{array}{c} 0.41 - 0.98 \\ 0.63 \pm 0.13 \end{array}$	$\begin{array}{c} 0.36-0.88\\ 0.57\pm 0.13\end{array}$	$\begin{array}{c} 0.56 - 0.82 \\ 0.65 \pm 014 \end{array}$.111
Ventral sucker maximum diameter (VSmax)	0.88 - 3.31 1.34 ± 0.33	$\frac{1.26 - 1.40}{1.33 \pm 0.09}$	1.26 - 2.36 1.70 ± 0.23	$\frac{1.14 - 2.96}{1.81 \pm 0.29}$	0.77 - 2.11 1.49 ± 0.36	1.22 - 1.84 1.61 ± 0.34	.000
Ventral sucker minimum diameter (VSmin)	0.81 - 2.31 1.22 ± 0.24	1.24 - 1.29 1.26 ± 0.03	$\begin{array}{c} 0.96 - 2.15 \\ 1.56 \pm 0.22 \end{array}$	0.82 - 2.54 1.66 ± 0.26	$\begin{array}{c} 0.63 - 2.06 \\ 1.35 \pm 0.37 \end{array}$	$\frac{1.06 - 1.77}{1.49 \pm 0.38}$.000

Discussion

The main goal of the current research was to identify Fasciola species by morphological methods. Various methods are used to determine the species of Fasciola^{4,20}. One of the most important methods for detection *Fasciola* is based on morphological differences in species^{5,10,18} Valero (2001) used *Fasciola hepatica* found in Altiplano in Bolivia as a

standard representatives of this species and Bargues (2005) used samples from Burkina Faso as standard representatives of *F. gigantica* because *Radix natalensis* is the only lymnaeid species (no *Galba/Fossaria*) in that country^{21,22}. In the past two decades researchers modified morphological methods were used to identify the species *Fasciola* and this modified method called computer image analysis system (CIAS)^{5,8,9}. In the current study, CIAS methods

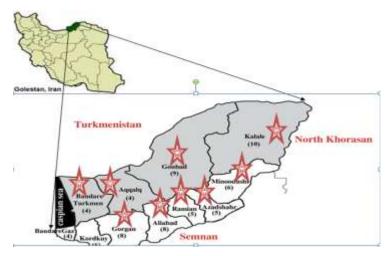


Figure 1. It shows Golestan province and locations where the samples were collected.

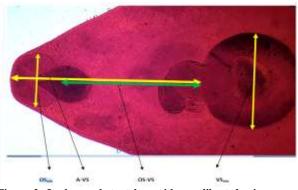


Figure 2. It shows photo taken with a calibrated microscope (Leica Galen III).

were used for detection of *Fasciola* species. Periago et al (2006) using (CIAS) succeeded in identifying *F. gigantica* from *F.hepatica* in both Europe and Africa²³. The results of this research are shown in Table 1. Most species isolated from Golestan province's was *Fasciola gigantica* (51.31%) Table 1 shows the measured factors of *Fasciola* worms in Golestan province, the analysis of which was performed using SPSS software. According to the table, the average body length of *F. hepatica* worms in cattle is 17.6 mm but in sheep 26.1 mm. In addition, the average body length of *F. gigantica* in cow is 34.28 mm but in sheep 30.11 mm.

There are significant differences in the body length of parasites. The average Body width of *F. hepatica* worms in cattle is 7.6 mm but in sheep 12.2 mm. Ashrafi et al. (2015) conducted a study in the province of Guilan and found out that the dominant species on the plains and low altitudes was

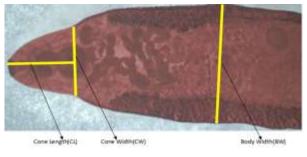


Figure 3. It shows photo taken with a calibrated stereomicroscope (Optica).

F.gigantica and at higher altitudes, the species was F. $hepatica^{24}$. Given that towns in the Golestan province were in the low-lying plains, F. gigantica isolated in the region is more consistent with the Ashrafi's study. It was also indicated that most sheep in Golestan were infected with F. hepatica. On the other hand, most cattle were infected with F. gigantica that is in line with Rokni et al.'s study $(2010)^{25}$. The findings of the present study showed that 36.84% from Fasciola was of F. hepatica species. Therefore, two species of Fasciola were found in Golestan province. Periago et al. (2006) believe that if two species of F. hepatica and F. gigantica coexist in an area, the intermediate form will be reported as Fasciola sp. Based on this study and according to Periago et al.'s research report 27 out of 228 samples investigated in the current study were not identified and were equivalent to 11.84% of all sample worms indicated as Fasciola sp. In this study, 88.16% of worms were identified by morphological methods. Although this method is time-

Species City		F.hepatica	F.£	rigantica	F	<i>Fasciola</i> .sp	Т	`otal
Gorgan	5	10.2%	33	67.3%	11	22.4%	49	100%
Gonbad-e kavus	5	14.28%	27	77.1%	3	8.57%	35	100%
Aliabad	18	54.54%	12	36.36%	3	9.09%	33	100%
Ramian	10	50%	10	50%	-	-	20	100%
Azadshaher	13	52%	9	36%	3	12%	25	100%
Bandar turkman	17	68%	4	16%	4	16%	25	100%
Aq Qala	14	87.5%	-	-	2	12.5%	16	100%
Minudesht	-	-	18	100%	-	-	18	100%
Kalaleh	2	28.57%	4	57.14%	1	14.28%	7	100%
Total	84		117		27		228	

Table 2: List of towns under investigation and separated species of Fasciola.

Table 3: Species identified based on host in Golestan province.

		Cattle				Sheep	
Total	Fasciola sp	F. hepatica	F. hepatica	Total	Fsciola sp	F. gigantica	F. hepatica
83	3	78	2	145	14	39	82
100%	3.61%	93.97%	2.4%	100%	16.55%	26.89%	56.55%

consuming and cumbersome in nature, morphological identification method is an appropriate way to identify *Fasciola gigantica* from *Fasciola hepatica*.

Conclusion

This study indicated that the main two species of worms, that is *F. hepatica* and *F. gigantica* were found in abundance in Golestan province. These two species of *Fasciola* in the area under investigation were found in overlapping coexistence. The current study was unable to identify 11.84% genus *Fasciola* showed as *Fasciola sp*.

Acknowledgment

The paper is taken from Ph.D. dissertation of Ahmad Halakou's, International Branch Shahid Beheshti University of Medical Sciences and research grant number 5139 supported by School of Medicine at Shahid Beheshti University of Medical Sciences,Tehran- Iran. The authors would like to thank the Veterinary Organization of Golestan Province for assistance in obtaining infeceted liver and fluke parasite samples.

References

1. Mas-Coma S, Bargues M D. Human liver flukes: A review. Research and Reviews in Parasitology.1997; 57: 145–218.

2. Boray J C. Fasciolosis. In: Hillyer GV, Hopla CE (Eds) Handbook Series in Zoonoses, Vol. III. CRC, Boca Raton- Florida. 1982;pp 71– 88.

3. Chen M G & Mott K E. Progress in assessment of morbidity due to Fasciola hepatica infection: a review of recent literature. Tropical diseases bulletin. 1990; 87(4): 1-38.

4. Mas-coma S, Valero M A, Bargues MD. Chapter2 Fasciola, Lymnaeids and human fascioliasis with a global overview on disease transmission, epidemiology, evolutionary genetics, molecular epidemiology and control. Advances in Parasitology. 2009; 69:41-146

5. Ashrafi K , Valero M A , Panova M , Periago M V, Massoud J, Mas-Coma S. Phenotypic analysis of adults of Fasciola hepatica, Fasciola gigantica and intermediate forms from the endemic region of Gilan, Iran. Parasitology International. 2006; 55: 249–260

6. Ashrafi K, Massoud J, Holakouie K, Jo-Afshani MA, Mahmoodi M, Ebadati N, Rezvani, Artigas P, Bargues MD, Mas-coma S. Nuclear ribosomal DNA ITS-2 sequence characterization of Fasciola hepatica and Galba trancatula. Iranian Journal of Public Health. 2007 ; 36(4): 42-49

7. Gradwohl, S. Clinical Laboratory Methods and Diagnosis, 7th Edition. The CV Mosby Company, St. Louis, Toronto, London. 1970; p: 1764

8. Valero MA, Panova M, Mas-Coma S. Development differences in the uterus of Fasciola hepatica between livestock liver fluke populations from Bolivian highland and European lowlands Parasitology Research. 2001; 87: 337–342 9. Dar Y, Vignoles P, Dreyfuss G, Rondelaud D. Fasciola hepatica and Fasciola gigantica: comparative morphometric studies on the redial stage of both species. Parasitology Research. 2003; 91: 369–373

10. Lotfy W M, El-Morshedy H N, Abou El-Hoda M, El-Tawila, Omar E A , Farag HF. Identification of the Egptian species of *Fasciola*. Veterinary Parasitology. 2002; 103: 323-332

11. Assmar M, Milaninia A, Amirkhani A, Yadegari D, Forghanparast K, Nahravanian H, Piazak N, Esmayli A, Hovanesian A, Aj Valadkhani Za. Seroepidemiological investigation of fascioliasis in northern Iran. Medical Journal of the Islamic Republic of Iran. 1991; 15;5(1):23-7.

12. Hosseini S H, Vaezi V, Jafari G, Rezaei A, Carami G. Epidemiological study of Fasciolosis in Kermanshah Province. Journal of Veterinary Faculty, University of Tehran. 2004; 59(3) : 201-206

 Salahimoghaddam A. Epidemiology of Human Fascioliasis in Iran. Journal of Kerman University of Medical Science, 2009; 16(4):385-394

14. Hatami H, Asmar M, Masoud J, Mansouri F, Namdaritabar H and Ramazankhani A. The first epidemic and new-emerging human fascioliasis in Kermanshah(western iran) and ten-year follow up, 1998-2008. International Journal of Preventive Medicine. 2012; 3(4):266-272.

15. Ashrafi K, Bargues MD, O'Neill S, Mas-Coma S. Fascioliasis: A worldwide parasitic disease of importance in travel medicine. Travel medicine and infectious disease. 2014 Dec 31;12(6):636-49.

16.Eslami A. Veterinary Parasitology, Tehran University Press. 2006; pp: 840-844.

17. Valero MA, Marcos MD, Mas-Coma S-A. Mathematical model for the ontogeny of Fasciola hepatica in the definitive host. Research and Reviews in Parasitology.1996; 56:13–20.

18. Valero M A, Panova M, Mas-Coma S. Phenotypic analysis of

adults and eggs of Fasciola hepatica by computer image analysis system. Journal of Helminthology. 2005; 79: 217–225.

19. Periago M V, Valero M A, El Sayed M, Ashrafi K, El Wakeel A, Mohamed MY, Desquesnes M, Curtale F, Mas-Coma S. First phenotypic description of Fasciola hepatica/Fasciola gigantica intermediate forms from the human endemic area of the Nile Delta, Egypt. Infection, Genetics and Evolution. 2008;8: 51–58.

20. Mas-Coma, S. Epidemiology of fascioliasis in human endemic areas. Journal of Helminthology. 2005; 79: 207–216.

21. Valero M A, Darce N A, Panova M, Mas-Coma S. Relationships between host species and morphometric patterns in Fasciola hepatica adults and eggs from the Northern Bolivian Altiplano hyperendemic region. Veterinary Parasitology. 2001a:102: 85–100.

22. Bargues MD, Mas-Coma S . Reviewing lymnaeid vectors of fasciolosis by ribosomal DNA sequence analyses. Journal of Helminthology. 2001a;79:257–267

23. Periago M V, Valero M A, Panova M, Mas-Coma S. Phenotypic comparison of allopatric populations of Fasciola hepatica and Fasciola gigantica from European and African bovines using a computer image analysis system (CIAS). Parasitology Research. 2006; 99: 368–378.

24. Ashrafi K, Valero M A, Peixoto R V, Artigas P, Panova M, Mas-Coma S. Distribution of Fasciola hepatica and F. gigantica in the endemic area of Guilan, Iran: Relationships between zonal overlap and phenotypic traits. Infection, Genetics and Evolution. 2015; 31 :95–109.

25. Rokni MB, Mirhendi H, Mizani A, Mohebali M, Sharbatkhori M, Kia EB, Abdoli H, Izadi S. Identification and differentiation of Fasciola hepatica and Fasciola gigantica using a simple PCR-restriction enzyme method. Experimental parasitology. 2010 Feb 28;124(2):209-13.