Review Article

Histopathologic Changes and Cellular Events of Organs Systems in COVID-19

Niloufar Najar Nobari¹¹, Fatemeh Montazer², Farnoosh Seirafianpour³, Farahnaz Nikkhah⁴, Zeinab Aryanian^{5,6}, Azadeh Goodarzi^{1*}

Abstract

A better understanding of histopathologic and cellular events of COVID-19 can help us to choose more proper management strategies and treatments. There are several ways to cellular and histopathologic evaluations; including, tissue sampling from living people as biopsies or dead people as autopsies or necropsies. In this study, we tried to evaluate the histopathologic concordance between findings of various tissue samplings derived from different sites that may work as a mirror of each other, especially mucocutaneous findings which may be indicative of similar events of other parts. Based on the main keywords, we searched databases of PubMed, Scopus, Google Scholar, Medscape, and CEBD coronavirus dermatology resource of Nottingham University and included the most relevant and well-designed studies with a higher level of pieces of evidence and higher sample size. In this study, selected pathological samples from different tissues of patients with COVID-19, including skin and mucosa, lungs, gastrointestinal tract, and kidneys were studied and summarized. Cellular changes and pathological findings in these patients were included by organ and listed by prevalence. Tissue sampling in patients with COVID-19 may help understand the pathophysiology of the disease as much as possible. Although most of these samples are taken after dying the patients, sampling before the more advanced stages of the disease could also show signs of tissue involvement before appearing the full systemic symptoms.

Keywords: Corona, COVID-19, Biopsy, Autopsy, Necropsy, Tissue sampling, Histopathology, Immunohistochemistry (IHC), Histology, Pathology

1. Department of Dermatology, Rasool Akram Medical Complex, Iran University of Medical Sciences (IUMS), Tehran, Iran

2. Department of Pathology, Firoozabadi Clinical Research Development Unit (FCRDU), Iran University of Medical Sciences (IUMS), Tehran, Iran

 Student Research Committee, School of Medicine, Iran University of Medical Sciences (IUMS), Tehran, Iran
Rasool Akram Medical Complex Clinical Research Development Center (RCRDC), Iran University of Medical Sciences, Tehran, Iran

 Autoimmune Bullous Diseases Research Center, Tehran University of Medical Sciences, Tehran, Iran
Department of Dermatology, Babol

University of Medical Sciences, Babol, Mazandaran, Iran

Corresponding Author:

Dr. Azadeh Goodarzi, Department of Dermatology, Rasool Akram Medical Complex, Niayesh St. Sattarkhan Ave., Tehran, Iran. E mail: Azadeh_goodarzi1984@yahoo.com

Please cite this article as: Najar Nobari N, Montazer F, Seirafianpour F, Nikkhah F, Aryanian Z, Goodarzi, A. Histopathologic Changes and Cellular Events of Organs Systems in COVID-19. J Cell Mol Anesth. 2021;6(1):81-88. DOI: https://doi.org/10.22037/jcma.v6i1.32528

Introduction

In the pandemic era of COVID-19, it is important to know what happens in the disease course in different parts of our body. Involvement of each site could be a presenting sign of a new coronavirus with its special clinical symptoms, but "how we can access the real data in this regard?"

Tissue sampling evaluations of different organsin living people as biopsies or in dead people as autopsies- give access to histopathological events during the COVID-19 course. To further improve our knowledge about the probable underlying pathophysiology of affected tissues and taking a better approach to the treatment, it is needed to understand more about histopathological events. In COVID-19, skin biopsy of affected patients and primary viralrelated cutaneous manifestation could be an appropriate diagnostic method; however, a biopsy of internal organs can be done under certain conditions but it is very rarely necessary and complex, and accompanied by legal ethical issues, while the autopsy of internal organs and their histopathological examination is much easier under the observance of ethical and legal rules that it is quite possible and acceptable.

Based on autopsies, COVID-19 is a systemic disease with major cardiovascular and pulmonary involvement in a cytokine storm and inflammatory setting, associated with a hypercoagulative state and susceptibility to microangiopathy and micro/macro thrombotic events (1).

Many efforts have been done to prepare autopsy guidelines per detail. Here, we would like to answer these questions regarding the COVID-19 course:

- Is skin biopsy somehow indicative or predictive of histopathological events of internal organs so to be considered as a substitute for internal organ biopsy, necropsy, or autopsy?
- Is skin biopsy indicative or predictive of histopathological events of internal organs? Is there histopathological concordance?
- Can tissue samples and pathological findings predict the course of the disease or the occurrence of expected clinical manifestations?

In the setting of pathologic concordance between data of various tissue samplings, perhaps the maximum biopsy of all available cases with affected skin is a good reason to raise our knowledge regarding the pathogenesis of the disorder. This is what we tried to approach in this comprehensive review.

We searched databases of PubMed, Scopus, Google Scholar, Medscape, and CEBD coronavirus dermatology resource of Nottingham University (https://www.nottingham.ac.uk/). Our keywords were ("COVID-19") OR ("severe acute respiratory syndrome coronavirus2") AND ("Skin") OR ("Cutaneous") OR ("Mucosa") OR ("Appendageal") OR ("Skin Manifestations") OR ("Dermatology") OR

("Histology") ("Pathology") OR OR ("Histopathology") OR ("Histopathologic") OR ("Histology") OR ("Biopsy") OR ("Autopsy") OR ("Necropsy"). For writing this comprehensive review, we included the most relevant, large (higher sample size) and well-designed studies with a higher level of evidence. The date of our search was from January 2020 to June 2020. We searched these databases for articles including data of patients who died from COVID-19 and being undergone autopsies for histopathological evaluation of different vital internal organs especially the lung, heart, and liver. In our search, we also found a living patient who was assessed after lung transplantation for histopathologic changes of the extracted lungs. We tried to discuss pathological events in different parts of the dead people and compare the histopathological findings between various organs even mucocutaneous changes. We focused on similarities or discrepancies for future better judgments about real COVID-19 pathogeneses as well as the events that may lead to different organ failures or death.

In this review, we focused on some main categories of histopathologic evaluations in COVID-19 patients; such as mucocutaneous and skin appendage histopathologic findings primarily related to the virus itself (not to its therapy (adverse cutaneous drug reactions)), pulmonary and other internal organ tissue sample or biopsy's histopathologic data and also the histopathologic changes and features in various sites of autopsies (2-20). The findings of the articles are reviewed and the results are presented in the order of prevalence in the table below (Table 1).

Novel coronavirus pneumonia caused by SARS-CoV-2 has a worldwide outbreak since it firstly occurred in December 2019, Wuhan city, China (21, 22). It was reported that the virus homology was over 85% between novel coronavirus pneumonia and severe acute respiratory syndrome (SARS). It means that the pathological changes of COVID-19 might be similar to SARS patients (23). In various tissue samples taken from patients with COVID-19, various manifestations can be observed depending on the involvement of the tissue and clinical symptoms. Using histology in these patients may raise our understanding of the pathophysiology of the COVID-19 virus. Histological studies in these patients mostly include

. . .

Mucocutaneous	Sub corneal pustules
pathology	• Spongiosis
	Papillary dermal edema
	• Perivascular and interstitial inflammatory cells infiltration consist of neutrophil and eosinophil
	Erythrocyte extravasation
	Microthrombi /fibrin thrombi
	• Vasculitis (usually necrotizing rather than lymphocytic)
	• Necrosis of the epidermis and dermis
	Vacuolar degeneration of the basal layer
	Lichenoid dermatitis
	Lymphocytic exocytosis to the epidermis and acrosyringia
	Dyskeratotic cells/ ballooning multinucleated keratinocytes
	Superficial vascular ectasia
	(Blue: epidermal changes, Green: dermal changes)
Pulmonary	Bilateral diffuse alveolar damage (DAD)
pathology	Proteinaceous exudate
	Pneumocyte hyperplasia
	Hyaline membrane formation
	Syncytial giant cell forms
	Interstitial fibrosis
	Vascular lumen stenosis
	Micro thrombosis formation
	Alveolar cavity congestion
	Desquamation and squamous metaplasia in alveolar epithelial cells
	Inclusion of viral bodies
Cardiac	Mild lymphocytic myocarditis
Pathology	• Epicarditis
	Focal mild fibrosis
	Mild myocardial hypertrophy
	• Interstitial fibrosis
Liver Pathology	Periportallymphoplasmacytic infiltration
	Micro vesicular steatosis
	Centrilobular sinusoidal dilation

Table 1: COVID-19-induced histopathologic changes in biopsy or autopsy samples of different sites.

	Patchy hepatic necrosis
	Fibrosis
	Nuclear glycogenetic in hepatocytes
	A mild increase in sinusoidal lymphocytes
	Kupffer cells hyperplasia in few sinusoidal spaces
Splenic pathology	Diminished white pulp with loss of marginal zones
	• Expansion of red pulp with lymphoplasmacytic infiltrate
Placental	• Thrombosis
pathology	Intramural fibrin deposition
	Villous stromal-vascular karyorrhexis
	• High-grade chronic villitis with associated avascular villi (obliterative vasculopathy) impaired vascular perfusion
	• Decidualvasculopathy
	Acute chorioamnionitis and funisitis
	Meconium-stained amniotic fluid and meconium macrophages
	Basal chronic villitis
	Hypercoiled umbilical cord, distal villous immaturity
	• Funisitis
	Retroplacental hematoma
	Cord hemangioma
Renal pathology	Loss of tubular brush border
	• Vacuolar degeneration of the tubular epithelium
	• Dilatation of the tubular lumen with cellular debris
	Necrosis and detachment of tubular epithelium
	• Hemosiderin granules in tubular epithelium
	Lymphocytic infiltration subcapsular areas
	Pseudo crescent appearance in glomerular structure

mucocutaneous, lung, liver, and kidney biopsies. Many of these findings have been derived from post-mortem sampling. Skin manifestations are also abundant in these patients, ranging from viral exanthema to vascular manifestations. Skin lesions may occur at any stage of the disease from asymptomatic carriers to patients admitted to the intensive care unit (ICU). Biopsy of these lesions shows different cases that correspond to the bed, such as Microthrombi/ fibrin thrombi, vasculitis, necrosis of the epidermis and dermis, and lichenoid changes. The lungs of the patients involved with COVID-19 showed diffuse alveolar damage and infiltrating perivascular lymphocytes with diffuse congestion and hemorrhagic necrosis, proliferative bronchiolitis, and alveolitis with pneumocytes proliferation, alveolar epithelial atrophy, epithelial desquamation, and squamous metaplasia. Similar changes in association with vascular damages include various degrees of inflammatory reactions and host immune reaction with variable tissue damages, like those seen in skin and lung biopsies, that all may be observed in other organs, such as the liver and kidneys. Examination of the pathological changes in the tissue can predict the manifestations of the disease and its course (24). Although most of these tissue samples especially from the lungs, liver, or kidneys have been prepared since the patient's death, evidence of the tissue involvement can be effective in the judgment of the disease.

Mucocutaneous findings are prevalent in COVID patients and studies report that about 20% of the patients have viral associated dermatologic signs that most of them are clinically compatible with maculopapular exanthematous or urticarial rash and vasculopathy or vasculitic features specially COVID Toes (8, 25-28). Histology findings along with viral affected pathologic changes and nonspecific inflammatory reactions indicate that an important group of patients have thrombotic microvascular events that are related to certain clinical presentations and laboratory hypercoagulative states in some patients (12, 29, 30).

A whole-lung biopsy of a patient who developed acute respiratory distress syndrome (ARDS) and respiratory failure and underwent lung transplantation showed diffuse congestion some hemorrhagic necrosis in the tissue. Gross examination and serial cuts microscopic histopathologic evaluation showed proliferative bronchiolitis and alveolitis with Pneumocytes hyperplasia, atrophy of alveolar epithelium, epithelial desquamation, and squamous metaplasia as well as massive fibrosis with hyaline degeneration and various stages of hemorrhagic infarction. Massive fibrosis is confirmed by Masson staining. The proliferation of thickened wall small vessels, luminal stenosis, occlusion, and microthrombosis was another main finding. Monocytes, lymphocytes, and plasma cells were infiltrated focally to the lung. Infiltrating into the pulmonary interstitium, and Immunohistochemistry (IHC) showed CD3, CD4, CD8, CD20, CD79a, CD5, CD38, and CD68 positive immune cells. Alveolar epithelial cells showed degeneration, atrophy, vacuolar proliferation, desquamation, and squamous metaplasia, and the alveolar cavity was congested and filled by mucus,

edema fluid, desquamated epithelial cells, and various degrees of inflammatory cells. Multinucleate giant cells and intracytoplasmic and intranuclear viral inclusion bodies (as viral cytopathic effect) also were found. In the COVID-19 course, several representative cytokines have been identified including IL-1 β , IL-18, TNF- α , IL-6, IL-8, and IL-10, which are produced and regulated by various immunological cells including CD8 and CD4 T cells (31).

There are many articles regarding autopsies of internal organs in COVID-19 patients, especially pulmonary, cardiac, and liver samples. In the lung, cytopathic effects usually with no obvious intranuclear or intracytoplasmic viral inclusions could also be identified as multinucleated syncytial cells with atypically enlarged pneumocytes characterized by large nuclei, amphophilic granular cytoplasm, and prominent nucleoli in intra-alveolar spaces. Fibrotic pathogenic events during systemic involvements in COVID-19 may be seen, especially in the lung. Moreover, important proteinaceous exudates as large protein globules, vascular congestion accompanied by inflammatory clusters of multinucleated fibrinoid materials, giant cells; pneumocyte hyperplasia and bilateral diffuse alveolar damage (DAD) with cellular fibromyxoid exudates also may be observed. Also, desquamated pneumocytes and hyaline membrane formation, suggesting interstitial mononuclear inflammatory infiltrates, were reported in ARDS dominated by lymphocytes (32, 33).

In postmortem needle core biopsies of the lung, liver, and heart of four patients who died of COVID-19 pneumonia, the main histopathologic finding of the lung was diffuse alveolar damage (DAD), including injury to the alveolar epithelial cells, hyaline membrane formation, and pneumocytes type II proliferation. Fibrotic degeneration consists of fibroblastic proliferation with the extracellular matrix, and fibrin cluster formation in airspaces was found. In the case of high intra-alveolar neutrophilic infiltration, there was bacterial superinfection. Changes in the liver and heart are likely secondary or related to the underlying diseases or are drug-dependent (especially in liver histopathological changes). The liver exhibited mild lobular infiltration by small lymphocytes, and centrilobular sinusoidal dilation along with patchy

necrosis, and the heart showed only focal mild fibrosis and mild myocardial hypertrophy (34).

The most important findings of recent studies indicate that the pulmonary small vessel and capillary thrombosis and microangiopathy with associated hemorrhage can be probably the main cause of the death. Severe endothelial injury associated with the presence of the intracellular virus and disrupted cell membranes and widespread thrombosis with microangiopathy could a characteristic feature of the lung involvement in COVID-19; so that alveolarcapillary microthrombi could be 9 times more prevalent than that in patients with influenza, and new vessel growth (intussusceptive angiogenesis) could be 2.7 times more prevalent than that in patients with influenza.

DAD including hyaline membrane formation could be seen in all patients, even non-intubated ones, and it is similar to histopathologic features of coronavirus as a severe acute respiratory syndrome (SARS). In the heart, individual cell necrosis without lymphocytic myocarditis is the most characteristic finding. Secondary pulmonary infection by microorganisms is of great concern nowadays. Central Nervous System did not show any involvement (2, 34, 35).

Although viral inclusion bodies in lung autopsies are not usual in an autopsy sample from Iran, we observed intra-alveolar hemorrhage, moderately interstitial inflammation composed of lymphocytes, plasma cells and neutrophils, focally necrosis and alveolar destruction, intravascular micro-thrombi, and large multinucleated pneumocytes with the focal presence of intranuclear inclusion like structure (maybe due to viral cytopathic effect), desquamation of pneumocytes without any evidence of mucus plugging within airways. Furthermore, in the liver autopsy of another case from Iran, we observed mild lobular and portal inflammation dominated by lymphocytes, moderate macrovesicular and mild microvesicular steatosis with diffuse distribution, moderate ballooning degeneration of hepatocytes with moderate bile pigment (36).

Due to the somewhat histopathological similarity between the involvements of different organs, it is important to know" which organ or organs are the main targets of the COVID-19 virus?"

Since most of the severe complications and deaths caused by the virus are due to lung involvement, so the lung can be the main target of the virus and other organs may be involved secondarily; however, they could act as a good predictor of pathologic events in the main target organ. Skin is the best example with more evident and accessible data among these secondary organs (37).

However, some facts about the actual course of COVID-19, the sequels, prognostic factors, treatments, and the manner of different parts of the body involvement would become clearer in the future, and the field of dermatology is not an exception. Some concerns are of great importance and need further investigation, including The virus-related or drug-related mucocutaneous signs of COVID-19 (38), management of patients with a specific dermatologic disorder in the pandemic era (39), or approach to the elective therapies especially in the elderly patients (40) like cosmetic procedures, non-emergent surgeries, or some chronic insignificant medical skin disorders (41), and more focus might be given to teledermatology.

The field of COVID is of interest to the authors of this review, and they are interested in COVID-19 issues in various fields especially COVID-19 and dermatology and now they think this topic may a valuable concern to discuss (42-45). In the pandemic era, there are many concerns about prevention, early diagnosis, and treatment of patients with COVID-19 (46-48) so here we tried to focus on a hot topic that may help to increase our knowledge about probable pathomechanisms that results to design better management protocols.

Conclusion

This study doubles the importance of tissue sampling. First, it helps to know more about the pathophysiology of the disease and how different tissues are involved. Second, it may be effective in predicting the course of the disease and subsequent clinical manifestations.

Acknowledgment

The authors would like to thank Rasool Akram Medical Complex Clinical research development Center (RCRDC) for its technical and editorial assists.

Conflicts of Interest

The authors declare that there are no conflicts of interest.

References

1. Buja LM, Wolf DA, Zhao B, Akkanti B, McDonald M, Lelenwa L, et al. The emerging spectrum of cardiopulmonary pathology of the coronavirus disease 2019 (COVID-19): Report of 3 autopsies from Houston, Texas, and review of autopsy findings from other United States cities. Cardiovasc Pathol. 2020;48:107233.

2. Ackermann M, Verleden SE, Kuehnel M, Haverich A, Welte T, Laenger F, et al. Pulmonary Vascular Endothelialitis, Thrombosis, and Angiogenesis in Covid-19. N Engl J Med. 2020;383(2):120-8.

3. Schaller T, Hirschbühl K, Burkhardt K, Braun G, Trepel M, Märkl B, et al. Postmortem Examination of Patients With COVID-19. JAMA. 2020;323(24):2518-20.

4. Herrero-Moyano M, Capusan TM, Andreu-Barasoain M, Alcántara-González J, Ruano-Del Salado M, Sánchez-Largo Uceda ME, et al. A clinicopathological study of eight patients with COVID-19 pneumonia and a late-onset exanthema. J Eur Acad Dermatol Venereol. 2020;34(9):e460-e4.

5. Recalcati S, Barbagallo T, Frasin LA, Prestinari F, Cogliardi A, Provero MC, et al. Acral cutaneous lesions in the time of COVID-19. J Eur Acad Dermatol Venereol. 2020;34(8):e346-e7.

6. García-Gil MF, García García M, Monte Serrano J, Prieto-Torres L, Ara-Martín M. Acral purpuric lesions (erythema multiforme type) associated with thrombotic vasculopathy in a child during the COVID-19 pandemic. J Eur Acad Dermatol Venereol. 2020;34(9):e443-e5.

7. Suarez-Valle A, Fernandez-Nieto D, Diaz-Guimaraens B, Dominguez-Santas M, Carretero I, Perez-Garcia B. Acro-ischaemia in hospitalized COVID-19 patients. J Eur Acad Dermatol Venereol. 2020;34(9):e455-e7. 8. Ahouach B, Harent S, Ullmer A, Martres P, Bégon E, Blum L, et al. Cutaneous lesions in a patient with COVID-19: are they related? Br J Dermatol. 2020;183(2):e31.

9. de Masson A, Bouaziz JD, Sulimovic L, Cassius C, Jachiet M, Ionescu MA, et al. Chilblains is a common cutaneous finding during the COVID-19 pandemic: A retrospective nationwide study from France. J Am Acad Dermatol. 2020;83(2):667-70.

11. Colonna C, Monzani NA, Rocchi A, Gianotti R, Boggio F, Gelmetti C. Chilblain-like lesions in children following suspected COVID-19 infection. Pediatr Dermatol. 2020;37(3):437-40.

12. Gianotti R, Zerbi P, Dodiuk-Gad RP. Clinical and histopathological study of skin dermatoses in patients affected by COVID-19 infection in the Northern part of Italy. J Dermatol Sci. 2020;98(2):141-3.

13. Gianotti R, Veraldi S, Recalcati S, Cusini M, Ghislanzoni M, Boggio F, et al. Cutaneous Clinico-Pathological Findings in three COVID-19-Positive Patients Observed in the Metropolitan Area of Milan, Italy. Acta Derm Venereol. 2020;100(8):adv00124.

14. Cordoro KM, Reynolds SD, Wattier R, McCalmont TH. Clustered cases of acral perniosis: Clinical features, histopathology, and relationship to COVID-19. Pediatr Dermatol. 2020;37(3):419-23.

15. Robustelli Test E, Vezzoli P, Carugno A, Raponi F, Gianatti A, Rongioletti F, et al. Acute generalized exanthematous pustulosis with erythema multiforme-like lesions induced by Hydroxychloroquine in a woman with coronavirus disease 2019 (COVID-19). J Eur Acad Dermatol Venereol. 2020;34(9):e457-e9.

16. Zengarini C, Orioni G, Cascavilla A, Horna Solera C, Fulgaro C, Misciali C, et al. Histological pattern in COVID-19-induced viral rash. J Eur Acad Dermatol Venereol. 2020;34(9):e453-e4.

17. Fernandez-Nieto D, Ortega-Quijano D, Jimenez-Cauhe J, Burgos-

Blasco P, de Perosanz-Lobo D, Suarez-Valle A, et al. Clinical and histological characterization of vesicular COVID-19 rashes: a prospective study in a tertiary care hospital. Clin Exp Dermatol. 2020;45(7):872-5.

18. Hanley B, Lucas SB, Youd E, Swift B, Osborn M. Autopsy in suspected COVID-19 cases. J Clin Pathol. 2020;73(5):239-42.

19. Santurro A, Scopetti M, D'Errico S, Fineschi V. A technical report from the Italian SARS-CoV-2 outbreak. Postmortem sampling and autopsy investigation in cases of suspected or probable COVID-19. Forensic Sci Med Pathol. 2020;16(3):471-6.

20. Mao D, Zhou N, Zheng D, Yue J, Zhao Q, Luo B, et al. Guide to forensic pathology practice for death cases related to coronavirus disease 2019 (COVID-19) (Trial draft). Forensic Sci Res. 2020;5(1):1-7.

21. Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, et al. A Novel Coronavirus from Patients with Pneumonia in China, 2019. N Engl J Med. 2020;382(8):727-33.

22. Holshue ML, DeBolt C, Lindquist S, Lofy KH, Wiesman J, Bruce H, et al. First Case of 2019 Novel Coronavirus in the United States. N Engl J Med. 2020;382(10):929-36.

23. Nicholls JM, Poon LL, Lee KC, Ng WF, Lai ST, Leung CY, et al. Lung pathology of fatal severe acute respiratory syndrome. Lancet. 2003;361(9371):1773-8.

24. Buja LM, Barth RF, Krueger GR, Brodsky SV, Hunter RL. The Importance of the Autopsy in Medicine: Perspectives of Pathology Colleagues. Acad Pathol. 2019;6:2374289519834041.

25. Galván Casas C, Català A, Carretero Hernández G, Rodríguez-Jiménez P, Fernández-Nieto D, Rodríguez-Villa Lario A, et al. Classification of the cutaneous manifestations of COVID-19: a rapid prospective nationwide consensus study in Spain with 375 cases. Br J Dermatol. 2020;183(1):71-7.

26. Estébanez A, Pérez-Santiago L, Silva E, Guillen-Climent S, García-Vázquez A, Ramón MD. Cutaneous manifestations in COVID-19: a new contribution. J Eur Acad Dermatol Venereol. 2020;34(6):e250-e1.

27. Recalcati S. Cutaneous manifestations in COVID-19: a first perspective. J Eur Acad Dermatol Venereol. 2020;34(5):e212-e3.

29. Young S, Fernandez AP. Skin manifestations of COVID-19. Cleve Clin J Med. 2020.

1. Buja LM, Wolf DA, Zhao B, Akkanti B, McDonald M, Lelenwa L, et al. The emerging spectrum of cardiopulmonary pathology of the coronavirus disease 2019 (COVID-19): Report of 3 autopsies from Houston, Texas, and review of autopsy findings from other United States cities. Cardiovasc Pathol. 2020;48:107233.

2. Ackermann M, Verleden SE, Kuehnel M, Haverich A, Welte T, Laenger F, et al. Pulmonary Vascular Endothelialitis, Thrombosis, and Angiogenesis in Covid-19. N Engl J Med. 2020;383(2):120-8.

3. Schaller T, Hirschbühl K, Burkhardt K, Braun G, Trepel M, Märkl B, et al. Postmortem Examination of Patients With COVID-19. JAMA. 2020;323(24):2518-20.

4. Herrero-Moyano M, Capusan TM, Andreu-Barasoain M, Alcántara-González J, Ruano-Del Salado M, Sánchez-Largo Uceda ME, et al. A clinicopathological study of eight patients with COVID-19 pneumonia and a late-onset exanthema. J Eur Acad Dermatol Venereol. 2020;34(9):e460-e4.

5. Recalcati S, Barbagallo T, Frasin LA, Prestinari F, Cogliardi A, Provero MC, et al. Acral cutaneous lesions in the time of COVID-19. J Eur Acad Dermatol Venereol. 2020;34(8):e346-e7.

6. García-Gil MF, García García M, Monte Serrano J, Prieto-Torres L, Ara-Martín M. Acral purpuric lesions (erythema multiforme type) associated with thrombotic vasculopathy in a child during the COVID-19 pandemic. J Eur Acad Dermatol Venereol. 2020;34(9):e443-e5.

7. Suarez-Valle A, Fernandez-Nieto D, Diaz-Guimaraens B, Dominguez-Santas M, Carretero I, Perez-Garcia B. Acro-ischaemia in hospitalized COVID-19 patients. J Eur Acad Dermatol Venereol. 2020;34(9):e455-e7. 8. Ahouach B, Harent S, Ullmer A, Martres P, Bégon E, Blum L, et al. Cutaneous lesions in a patient with COVID-19: are they related? Br J Dermatol. 2020;183(2):e31.

9. de Masson A, Bouaziz JD, Sulimovic L, Cassius C, Jachiet M, Ionescu MA, et al. Chilblains is a common cutaneous finding during the COVID-19 pandemic: A retrospective nationwide study from France. J Am Acad Dermatol. 2020;83(2):667-70.

10. Andina D, Noguera-Morel L, Bascuas-Arribas M, Gaitero-Tristán J,

Alonso-Cadenas JA, Escalada-Pellitero S, et al. Chilblains in children in the setting of COVID-19 pandemic. Pediatr Dermatol. 2020;37(3):406-11. 11. Colonna C, Monzani NA, Rocchi A, Gianotti R, Boggio F, Gelmetti

C. Chilblain-like lesions in children following suspected COVID-19 infection. Pediatr Dermatol. 2020;37(3):437-40.

12. Gianotti R, Zerbi P, Dodiuk-Gad RP. Clinical and histopathological study of skin dermatoses in patients affected by COVID-19 infection in the Northern part of Italy. J Dermatol Sci. 2020;98(2):141-3.

13. Gianotti R, Veraldi S, Recalcati S, Cusini M, Ghislanzoni M, Boggio F, et al. Cutaneous Clinico-Pathological Findings in three COVID-19-Positive Patients Observed in the Metropolitan Area of Milan, Italy. Acta Derm Venereol. 2020;100(8):adv00124.

14. Cordoro KM, Reynolds SD, Wattier R, McCalmont TH. Clustered cases of acral perniosis: Clinical features, histopathology, and relationship to COVID-19. Pediatr Dermatol. 2020;37(3):419-23.

15. Robustelli Test E, Vezzoli P, Carugno A, Raponi F, Gianatti A, Rongioletti F, et al. Acute generalized exanthematous pustulosis with erythema multiforme-like lesions induced by Hydroxychloroquine in a woman with coronavirus disease 2019 (COVID-19). J Eur Acad Dermatol Venereol. 2020;34(9):e457-e9.

16. Zengarini C, Orioni G, Cascavilla A, Horna Solera C, Fulgaro C, Misciali C, et al. Histological pattern in COVID-19-induced viral rash. J Eur Acad Dermatol Venereol. 2020;34(9):e453-e4.

17. Fernandez-Nieto D, Ortega-Quijano D, Jimenez-Cauhe J, Burgos-Blasco P, de Perosanz-Lobo D, Suarez-Valle A, et al. Clinical and histological characterization of vesicular COVID-19 rashes: a prospective study in a tertiary care hospital. Clin Exp Dermatol. 2020;45(7):872-5.

18. Hanley B, Lucas SB, Youd E, Swift B, Osborn M. Autopsy in suspected COVID-19 cases. J Clin Pathol. 2020;73(5):239-42.

19. Santurro A, Scopetti M, D'Errico S, Fineschi V. A technical report from the Italian SARS-CoV-2 outbreak. Postmortem sampling and autopsy investigation in cases of suspected or probable COVID-19. Forensic Sci Med Pathol. 2020;16(3):471-6.

20. Mao D, Zhou N, Zheng D, Yue J, Zhao Q, Luo B, et al. Guide to forensic pathology practice for death cases related to coronavirus disease 2019 (COVID-19) (Trial draft). Forensic Sci Res. 2020;5(1):1-7.

21. Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, et al. A Novel Coronavirus from Patients with Pneumonia in China, 2019. N Engl J Med. 2020;382(8):727-33.

22. Holshue ML, DeBolt C, Lindquist S, Lofy KH, Wiesman J, Bruce H, et al. First Case of 2019 Novel Coronavirus in the United States. N Engl J Med. 2020;382(10):929-36.

23. Nicholls JM, Poon LL, Lee KC, Ng WF, Lai ST, Leung CY, et al. Lung pathology of fatal severe acute respiratory syndrome. Lancet. 2003;361(9371):1773-8.

24. Buja LM, Barth RF, Krueger GR, Brodsky SV, Hunter RL. The Importance of the Autopsy in Medicine: Perspectives of Pathology Colleagues. Acad Pathol. 2019;6:2374289519834041.

25. Galván Casas C, Català A, Carretero Hernández G, Rodríguez-Jiménez P, Fernández-Nieto D, Rodríguez-Villa Lario A, et al. Classification of the cutaneous manifestations of COVID-19: a rapid prospective nationwide consensus study in Spain with 375 cases. Br J Dermatol. 2020;183(1):71-7.

26. Estébanez A, Pérez-Santiago L, Silva E, Guillen-Climent S, García-Vázquez A, Ramón MD. Cutaneous manifestations in COVID-19: a new contribution. J Eur Acad Dermatol Venereol. 2020;34(6):e250-e1.

27. Recalcati S. Cutaneous manifestations in COVID-19: a first perspective. J Eur Acad Dermatol Venereol. 2020;34(5):e212-e3.

28. Hedou M, Carsuzaa F, Chary E, Hainaut E, Cazenave-Roblot F, Masson Regnault M. Comment on 'Cutaneous manifestations in COVID-19: a first perspective' by Recalcati S. J Eur Acad Dermatol Venereol. 2020;34(7):e299-e300.

29. Young S, Fernandez AP. Skin manifestations of COVID-19. Cleve Clin J Med. 2020.

30. Magro C, Mulvey JJ, Berlin D, Nuovo G, Salvatore S, Harp J, et al.

Complement associated microvascular injury and thrombosis in the pathogenesis of severe COVID-19 infection: A report of five cases. Transl Res. 2020;220:1-13.

31. Shanmugam C, Mohammed AR, Ravuri S, Luthra V, Rajagopal N, Karre S. COVID-2019 - A comprehensive pathology insight. Pathol Res Pract. 2020;216(10):153222.

32. Xu Z, Shi L, Wang Y, Zhang J, Huang L, Zhang C, et al. Pathological findings of COVID-19 associated with acute respiratory distress syndrome. Lancet Respir Med. 2020;8(4):420-2.

33. Cascella M, Rajnik M, Cuomo A, Dulebohn SC, Di Napoli R. Features, Evaluation, and Treatment of Coronavirus (COVID-19). StatPearls. Treasure Island (FL): StatPearls Publishing.; 2021.

34. Tian S, Xiong Y, Liu H, Niu L, Guo J, Liao M, et al. Pathological study of the 2019 novel coronavirus disease (COVID-19) through postmortem core biopsies. Mod Pathol. 2020;33(6):1007-14.

35. Fox SE, Akmatbekov A, Harbert JL, Li G, Quincy Brown J, Vander Heide RS. Pulmonary and cardiac pathology in African American patients with COVID-19: an autopsy series from New Orleans. Lancet Respir Med. 2020;8(7):681-6.

36. Maiese A, Manetti AC, La Russa R, Di Paolo M, Turillazzi E, Frati P, et al. Autopsy findings in COVID-19-related deaths: a literature review. Forensic Sci Med Pathol. 2020:1-18.

37. Seirafianpour F, Mozafarpoor S, Fattahi N, Sadeghzadeh-Bazargan A, Hanifiha M, Goodarzi A. Treatment of COVID-19 with pentoxifylline: Could it be a potential adjuvant therapy? Dermatol Ther. 2020;33(4):e13733.

38. Seirafianpour F, Sodagar S, Pour Mohammad A, Panahi P, Mozafarpoor S, Almasi S, et al. Cutaneous manifestations and considerations in COVID-19 pandemic: A systematic review. Dermatol Ther. 2020;33(6):e13986.

39. Najar Nobari N, Seirafianpour F, Mashayekhi F, Goodarzi A. A systematic review on treatment-related mucocutaneous reactions in COVID-19 patients. Dermatol Ther. 2021;34(1):e14662.

40. Mohamadi M, Goodarzi A, Aryannejad A, Fattahi N, Alizadeh-Khoei M, Miri S, et al. Geriatric challenges in the new coronavirus disease-19 (COVID-19) pandemic: A systematic review. Med J Islam Repub Iran. 2020;34:123.

41. Ehsani A, Noormohammadpour P, Goodarzi A, Mirshams Shahshahani M, Hejazi SP, Hosseini E, et al. Comparison of long-pulsed alexandrite laser and topical tretinoin-ammonium lactate in axillary acanthosis nigricans: A case series of patients in a before-after trial. Caspian J Intern Med. 2016;7(4):290-3.

42. Nobari NN, Goodarzi A. Patients with specific skin disorders who are affected by COVID-19: What do experiences say about management strategies? A systematic review. Dermatol Ther. 2020;33(6):e13867.

43. Sahebnasagh A, Saghafi F, Avan R, Khoshi A, Khataminia M, Safdari M, et al. The prophylaxis and treatment potential of supplements for COVID-19. Eur J Pharmacol. 2020;887:173530.

44. Khalil S, Bardawil T, Stephan C, Darwiche N, Abbas O, Kibbi AG, et al. Retinoids: a journey from the molecular structures and mechanisms of action to clinical uses in dermatology and adverse effects. J Dermatolog Treat. 2017;28(8):684-96.

45. Rizk JG, Kalantar-Zadeh K, Mehra MR, Lavie CJ, Rizk Y, Forthal DN. Pharmaco-Immunomodulatory Therapy in COVID-19. Drugs. 2020;80(13):1267-92.

46. Mahmoodpoor A. SARS, MERS and COVID-19, the story continues. J Cell Mol Anesth. 2020;5(2):57-8.

47. Janbabai G, Razavi S, Dabbagh A. How to Manage Perioperative Patient Flow during COVID-19 Pandemic: a Narrative Review. J Cell Mol Anesth. 2020;5(1):47-56.

48. Faghihi Langroudi T, Khazaei M. Common imaging patterns of COVID-19 on spiral chest CT scan: a diagnostic approach for pulmonary involvement in ICU patients. J Cell Mol Anesth. 2020;5(1):6-14.