

## Chronic constipation: no more idiopathic, but a true neuropathological entity

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### ABSTRACT

Chronic constipation is still considered a functional or idiopathic disorder. However, in recent years there is evidence that its pathophysiological grounds may be actually due to a complex system of abnormalities of the enteric nervous system of these patients. In particular, as reported in this review, the enteric glial cells seem to be constantly involved in constipated patients, suggesting that (at least some forms of) constipation should be considered as true neuro-gliopathies.

**Keywords:** Constipation, Enteric glial cells, Enteric neurons, Enteric nervous system

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### Introduction

In recent years, the improvement of technology and the increase in knowledge have shifted several strongly held paradigms. This is particularly true in gastroenterology, and specifically in the field of the so-called “functional” or “idiopathic” disease, where conditions thought for decades to be based mainly on alterations of visceral perception or aberrant psychosomatic mechanisms have, in fact, be reconducted to an organic basis (or, at the very least, have shown one or more demonstrable abnormalities).

This is particularly true, for instance, for irritable bowel syndrome (IBS), the prototype entity of functional gastrointestinal (GI) disorders, where low-grade inflammation of both mucosa

and myenteric plexus has been repeatedly demonstrated (1, 2). Thus, researchers have also investigated other functional/idiopathic GI disorders, and found that some organic ground is present, such as abnormal neurotransmission and myenteric plexitis in esophageal achalasia (3-5) and mucosal immune activation and mild eosinophilia in functional dyspepsia (6, 7).

Here we show evidence, based on our own and other authors' work, that chronic constipation should no more be considered as a functional/idiopathic GI disorder, but instead as a true enteric neuropathic abnormality.

### What is known about pathophysiology of chronic constipation?

Chronic constipation is a frequent disorder, characterized by reduction of bowel movements and/or defecatory difficulties (8) but probably represents a multifaceted entity that may be

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broadly classified in two main subgroups (i.e., slow transit constipation, STC, and obstructed defecation, OD) (9).

The pathophysiological grounds of constipation have been traditionally attributed to the dysfunction of anorectal (10-16) and/or colonic motility (17, 18), even though the basic mechanisms responsible for this dysfunction still remains quite obscure. However, there is some evidence that constipated patients may have qualitative and quantitative changes in the enteric nervous system (ENS) of the colon such as abnormal enteric neurochemistry (19-21), reduction of intraganglionic neurofilaments (22), myenteric plexus hypoganglionosis (23), and decreased colonic interstitial cells of Cajal (ICC) (24, 25), even though discrepancies have been reported in other studies (26, 27).

### Recent neuropathological findings in chronic constipation

In these last years, thanks to the availability of full-thickness surgical specimens from patients with different subtypes of chronic constipation, we were able to investigate in a more comprehensive way some abnormalities of the ENS present in these subjects.

One of the subtypes of constipation more often labeled as intractable and refractory to any form of medical treatment is STC (28, 29), a form that sometimes requires a surgical approach (30). In a study on surgically resected STC patients, we demonstrated that, together with an important and significant decrease of enteric neurons (due to increased apoptotic phenomena) and ICC compared to controls, these patients also displayed a significant decrease of enteric glial cells (EGC) in both the myenteric and the submucous plexus, a decrease not due to increased apoptotic phenomena (31). Since similar neuropathological abnormalities were found in

the terminal ileum of these patients, we suggested that the neuropathologic process might not be limited to the colon in these subjects (32).

In another investigation, conducted on the ENS of patients with OD (in whom the pathophysiologic mechanisms are thought to be different) undergoing surgery for refractory symptoms, we found that, compared to control, a significant reduction of the enteric neurons only in the submucosal plexus, whereas there was a loss of EGC in both the myenteric and the submucosal plexus. No differences between patients and controls were found concerning ICC and the number of apoptotic neurons (33).

Subsequently, we studied colonic specimens of patients with idiopathic megacolon and Chagas's disease, undergoing surgery for intractable constipation (often the only therapeutic option in these cases (34); in both conditions we found a decreased number of enteric neurons and of EGC (the latter being more accentuated in chagasic patients) (35). Interestingly, other authors have then reported that, in patients with chagasic megacolon, the destruction of the enteric neurons is preceded by the loss of EGC, leading to the hypothesis that this cell population might prevent colonic dilatation, protecting the ENS against the inflammatory process and neuronal destruction (36).

Chronic constipation is a frequent feature of diverticular disease of the colon (37), but its pathophysiologic basis still remain largely unknown (38). In particular, very few data are available concerning ENS abnormalities in this condition (39). However, we and other authors documented that these patients, compared to controls, display a normal number of myenteric and submucosal plexus neurons, whereas ICC and EGC are significantly decreased (40, 41).

## Unifying pathophysiologic findings: the importance of EGC

From the above considerations, based on our own and other authors' work, it may be inferred that there is a common factor that unifies several conditions characterized by constipation, factor represented by a loss of colonic EGC. This and other observations have recently spurred us and other investigators to consider this cell population as an important determinant for gastrointestinal motor activity (especially in the colon) (42-45) and a factor of paramount importance in the pathophysiology of constipation (46, 47).

How can EGC be involved in such instances?

EGC represent most of the cell population of enteric ganglia, with a ratio of 4:1 compared to neurons (48) and, in addition to have a mechanical support function due to the adherence to the surface of enteric ganglia and nerve by means of filaments of glial fibrillary acidic protein (49). Also, they display several other functions such as participation in neurotransmission (50-52) and promotion of synaptic communication in enteric neurons (53), in the maintenance of the homeostatic function of the enteric neurons (54, 55), in inflammatory processes and immune functions of the gut (56, 57), and in visceral pain (58).

## How EGC participate in the pathogenesis of constipation

It must be firstly taken into account that, notwithstanding the above considerations, most data on EGC function originate from animal studies, and human data (apart from those reported above) are very scarce. However, we feel that some considerations and hypotheses may be formulated.

For instance, the absence of trophic glial factor is responsible for the loss of myenteric neurons containing substance P (an excitatory transmitter),

which in turn causes decrease of intestinal transit (59), and the loss of EGC impairs motility and transit in the gut (60). Since EGC provides a feedback system for ICC to modulate slow wave activity, their decrease in constipated patients, in whom ICC are also often decreased or absent, might further impair the reduced pacemaker signals originating from these cells (61).

Thus, on the above grounds, we have hypothesized that the decrease/loss of EGC (together with other factors, such as the concomitant decrease or loss of enteric neurons, ICC, enteric neurotransmitters, variation of colonic mast cell population (46, 47, 62)) plays a pivotal role in the pathogenesis of constipation. A decrease/loss of EGC, through impairment of neurotrophic factors, might for instance be responsible (similarly to that documented in experimental animal models) for degeneration of the enteric neurons population. In this respect, it is worth noting that in patients with "idiopathic" constipation in addition to the loss of EGC is often documented the concomitant decrease of enteric neurons due increased apoptotic phenomena (31, 32, 63); on the other hand, in constipated patients with degenerative conditions of the central nervous system (for instance, Alzheimer's disease) no EGC loss has been reported, and the reduction of enteric neurons is not due to apoptotic phenomena (64).

How can a selective loss/decrease of EGC be explained? Interestingly, recent data suggest that an infectious agent might play a role, as shown by the localization and multiplication of *Mycobacterium avium* subspecies and by the expression of prion protein in these cells (65-67). Moreover, a role for intestinal flora in the plasticity of the ENS has been recently hypothesized, since the population of EGC may be modulated by altered luminal environment (68).

The decrease of EGC could also be due to an aging process (69, 70), even though such data in human beings are only available for enteric

neurons (71). Other damaging factors on the ENS, such as that hypothesized by the use of anthraquinone laxatives, have not been confirmed with modern immunohistochemical techniques (72). On the other hand, the demonstration of chromosomal abnormalities of enteric neurons and EGC in severely constipated patients requiring surgery suggests the presence of a genetic background possibly responsible for the loss of these elements (73).

### Concluding remarks

Perhaps it is time that EGC are viewed not simply as the glue of the gut, but as important and pivotal cells that synchronize the various elements of the ENS. With this respect, the demonstration that several entities characterized by constipation display a decrease or loss of EGC, should lead (of course, in addition to other abnormalities) to classify constipation not as a “functional” or “idiopathic” condition, but as a neuro-gliopathy of the gut (46, 47, 74). Further studies are needed to establish a more precise role for EGC in the pathophysiology of gut motility.

### References

1. Chey WD, Cash BD. Irritable bowel syndrome: update on colonic neuromuscular dysfunction and treatment. *Curr Gastroenterol Rep* 2006; 8: 273-81.
2. Ohman L, Simrén M. Pathogenesis of IBS: role of inflammation, immunity and neuroimmune interactions. *Nat Rev Gastroenterol Hepatol* 2010; 7: 163-73.
3. Sigala S, Missale G, Missale C, Villanacci V, Cestari R, Grigolato PG, Lojcono L, Spano PF. Different neurotransmitter systems are involved in the development of esophageal achalasia. *Life Sci* 1995; 56: 1311-20.
4. Bruley des Varannes S, Chevalier J, Pimont S, Le Neel JC, Klotz M, et al. Serum from achalasia patients alters neurochemical coding in the myenteric plexus and nitric oxide mediated motor response in normal human fundus. *Gut* 2006; 55: 319-32.
5. Villanacci V, Annese V, Cuttitta A, Fisogni S, Scaramuzzi G, De Santo E, et al. An immunohistochemical study of the myenteric plexus in idiopathic achalasia. *J Clin Gastroenterol* 2010; 44:407-10.
6. Kindt S, Tertychnyy A, de Hertogh G, Geboes K, Tack J. Intestinal immune activation in presumed post-infectious functional dyspepsia. *Neurogastroenterol Motil* 2009; 21: 832-e56
7. Walker MM, Salehian SS, Murray CE, Rajendran A, Hoare JM, Negus R, et al. Implications of eosinophilia in the normal duodenal biopsy – an association with allergy and functional dyspepsia. *Aliment Pharmacol Ther* 2010; 31: 1229-36.
8. Rao SS, Meduri K. What is necessary to diagnose constipation? *Best Pract Res Clin Gastroenterol* 2011; 25: 127-40.
9. Lembo A, Camilleri M. Chronic constipation. *N Engl J Med* 2003; 349: 1360-68.
10. Preston DM, Lennard-Jones JE. Anismus in chronic constipation. *Dig Dis Sci* 1985; 30: 413-18.
11. Bassotti G, Morelli A, Whitehead WE. Abnormal rectosigmoid myoelectric response to eating in patients with severe idiopathic constipation (slow-transit type). *Dis Colon Rectum* 1992; 35: 753-76.
12. Rao SS, Welcher KD, Leistikow BS. Obstructed defecation: a failure of rectoanal coordination. *Am J Gastroenterol* 1998; 93: 1042-50.
13. Faucheron JL, Dubreuil A. Rectal akinesia as a new cause of impaired defecation. *Dis Colon Rectum* 2000; 43: 1545-49.
14. Gosselink MJ, Hop WC, Schouten WR. Rectal compliance in females with obstructed defecation. *Dis Colon Rectum* 2001; 44: 971-77.
15. Sloots CE, Felt-Bersma RJ. Rectal sensorimotor characteristics in female patients with idiopathic constipation with or without paradoxical sphincter contraction. *Neurogastroenterol Motil* 2003; 15: 187-93.
16. Karlbom U, Lundin E, Graf W, Pahlman L. Anorectal physiology in relation to clinical subgroups of patients with severe constipation. *Colorectal Dis* 2004; 6: 343-49.
17. Bassotti G, Iantorno G, Fiorella S, Bustos-Fernandez L, Bilder CR. Colonic motility in man: features in normal subjects and in patients with chronic idiopathic constipation. *Am J Gastroenterol* 1999; 94: 1760-70.
18. Bassotti G, Crowell MD. Colon and rectum: normal function and clinical disorder. In: Schuster MM, Crowell MD, Koch KL, eds. *Schuster atlas of*

gastrointestinal motility in health and disease. 2<sup>nd</sup> edition. Hamilton, Canada: BC Decker Inc.; 2002. p.241-52.

19. Koch T, Carney JA, Go VL. Idiopathic chronic constipation is associated with decreased colonic vasoactive intestinal peptide. *Gastroenterology* 1988; 94: 300-10.

20. Sjolund K, Fasth S, Ekman R, Hulten L, Jiborn H, Nordgren S, et al. Neuropeptides in idiopathic chronic constipation (slow transit constipation). *Neurogastroenterol Mot* 1997; 9: 143-50.

21. Zhao RH, Baig MK, Mack J, Abramson S, Woodhouse S, Wexner SD. Altered serotonin immunoreactivities in the left colon of patients with colonic inertia. *Colorectal Dis* 2002; 4: 56-60.

22. Schouten WR, ten Kate FJ, de Graaf EJ, Gilberts EC, Simons JL, Kluck P. Visceral neuropathy in slow transit constipation: an immunohistochemical investigation with monoclonal antibodies against neurofilament. *Dis Colon Rectum* 1993; 36: 1112-17.

23. Wedel T, Roblick UJ, Ott V, Eggers R, Schiedeck TH, Krammer HJ, et al. Oligoneuronal hypoganglionosis in patients with idiopathic slow transit constipation. *Dis Colon Rectum* 2002; 45: 54-62.

24. Lyford GL, He CL, Soffer E, Hull TL, Strong SA, Senatore AJ, et al. Pan-colonic decrease in interstitial cells of Cajal in patients with slow transit constipation. *Gut* 2002; 51: 496-501.

25. Wedel T, Spiegler J, Soellner S, Roblick UJ, Schiedeck THK, Bruch HP, et al. Enteric nerves and interstitial cells of Cajal are altered in patients with slow transit constipation and megacolon. *Gastroenterology* 2002; 123: 1459-67.

26. Yu CS, Kim HC, Hong HK, Chung DH, Kim HJ, Kang GH, et al. Evaluation of myenteric ganglion cells and interstitial cells of Cajal in patients with chronic idiopathic constipation. *Int J Colorectal Dis* 2002; 17: 253-58.

27. Toman J, Turina M, Ray M, Petras RE, Stromberg AJ, Galandiuk S. Slow transit colon constipation is not related to the number of interstitial cells of Cajal. *Int J Colorectal Dis* 2006; 21: 527-32.

28. Bassotti G, Roberto GD, Sediari L, Morelli A. Toward a definition of colonic inertia. *World J Gastroenterol* 2004; 10: 2465-67.

29. Bharucha AE. Constipation. *Best Pract Res Clin Gastroenterol* 2007; 21: 709-31.

30. Frattini JC, Nogueras JJ. Slow transit constipation: a review of a colonic functional disorder. *Clin Colon Rectal Surg* 2008; 21: 146-52.

31. Bassotti G, Villanacci V, Maurer CA, Fisogni S, Di Fabio F, Cadei M, et al. The role of glial cells and apoptosis of enteric neurones in the neuropathology of intractable slow transit constipation. *Gut* 2006; 55: 41-46.

32. Bassotti G, Villanacci V, Cathomas G, Maurer CA, Fisogni S, Cadei M, et al. Enteric neuropathology of the terminal ileum in patients with intractable slow-transit constipation. *Hum Pathol* 2006; 37: 1252-58.

33. Bassotti G, Villanacci V, Nascimbeni R, Asteria CR, Fisogni S, Nesi G, et al. Colonic neuropathological aspects in patients with intractable constipation due to obstructed defecation. *Mod Pathol* 2007; 20: 367-74.

34. Autschbach F, Gassler N. Idiopathic megacolon. *Eur J Gastroenterol Hepatol* 2007; 19: 399-400.

35. Iantorno G, Bassotti G, Kogan Z, Lumi CM, Cabanne AM, Fisogni S, et al. The enteric nervous system in chagasic and idiopathic megacolon. *Am J Surg Pathol* 2007; 31: 460-68.

36. da Silveira AB, Freitas MA, de Oliveira EC, Neto SG, Luquetti AO, Furness JB, et al. Glial fibrillary acidic protein and S-100 colocalization in the enteroglia cells in dilated and nondilated portions of colon from chagasic patients. *Hum Pathol* 2009; 40: 244-51.

37. Hemming J, Floch M. Features and management of colonic diverticular disease. *Curr Gastroenterol Rep* 2010; 12:399-407.

38. Jeyarajah S, Papagrigoriadis S. Review article: the pathogenesis of diverticular disease--current perspectives on motility and neurotransmitters. *Aliment Pharmacol Ther* 2011; 33: 789-800.

39. Brian West A. The pathology of diverticulosis: classical concepts and mucosal changes in diverticula. *J Clin Gastroenterol* 2006; 40: S126-31.

40. Bassotti G, Battaglia E, Bellone G, Dughera L, Fisogni S, Zambelli C, et al. Interstitial cells of Cajal, enteric nerves, and glial cells in colonic diverticular disease. *J Clin Pathol* 2005; 58: 973-77.

41. Wedel T, Büsing V, Heinrichs G, Nohroudi K, Bruch HP, Roblick UJ, et al. Diverticular disease is associated with an enteric neuropathy as revealed by morphometric analysis. *Neurogastroenterol Motil* 2010; 22: 407-14.

42. Bassotti G, Villanacci V, Antonelli E, Morelli A, Salerni B. Enteric glial cells: new players in gastrointestinal motility? *Lab Invest* 2007; 87: 628-32.

43. Savidge TC, Sofroniew MV, Neunlist M. Starring roles for astroglia in barrier pathologies of gut and brain. *Lab Invest* 2007; 87: 731-36.
44. Bassotti G, Villanacci V, Fisogni S, Rossi E, Baronio P, Clerici C, et al. Enteric glial cells and their role in gastrointestinal motor abnormalities: introducing the neuro-gliopathies. *World J Gastroenterol* 2007; 13: 4035-41.
45. De Winter BY, De Man JG. Interplay between inflammation, immune system and neuronal pathways: effect on gastrointestinal motility. *World J Gastroenterol* 2010; 16: 5523-35.
46. Bassotti G, Villanacci V. Slow transit constipation: a functional disorder becomes an enteric neuropathy. *World J Gastroenterol* 2006; 12: 4609-13.
47. Bassotti G, Villanacci V. Can "functional" constipation be considered as a form of enteric neurogliopathy? *Glia* 2011; 59: 345-50.
48. Jessen KR. Glial cells. *Int J Biochem Cell Biol* 2004; 36: 1861-67.
49. Schemann M, Neunlist M. The human enteric nervous system. *Neurogastroenterol Motil* 2004; 16: 55-59.
50. Giaroni C, Zanetti E, Chiaravalli AM, Albarello L, Dominioni L, Capella C, et al. Evidence for a glutamatergic modulation of the cholinergic function in the human enteric nervous system via NMDA receptors. *Eur J Pharmacol* 2003; 476 :63-69.
51. Braun N, Sevigny J, Robson SC, Hammer K, Hanani M, Zimmermann H. Association of ecto-ATPase NTP-Dase2 with glial cells of the peripheral nervous system. *Glia* 2004; 45: 124-32.
52. Gulbransen BD, Sharkey KA. Purinergic neuron-to-glia signaling in the enteric nervous system. *Gastroenterology* 2009; 136: 1349-58.
53. Zeng F, Watson RP, Nash MS. Glial cell-derived neurotrophic factor enhances synaptic communication and 5-hydroxytryptamine 3a receptor expression in enteric neurons. *Gastroenterology* 2010; 138: 1491-501.
54. Aube AC, Cabarrocas J, Bauer J, Philippe D, Aubert P, Doulay F, et al. Changes in enteric neurone phenotype and intestinal functions in a transgenic mice model of enteric glia disruption. *Gut* 2006; 55: 630-37.
55. Abdo H, Derkinderen P, Gomes P, Chevalier J, Aubert P, Masson D, et al. Enteric glial cells protect neurons from oxidative stress in part via reduced glutathione. *FASEB J* 2010; 24: 1082-94.
56. Cabarrocas J, Savidge TC, Liblau RS. Role of enteric glial cells in inflammatory bowel disease. *Glia* 2003; 41: 81-93.
57. von Boyen GB, Steinkamp M, Geerling I, Reinshagen M, Schäfer KH, Adler G, et al. Proinflammatory cytokines induce neurotrophic factor expression in enteric glia: a key to the regulation of epithelial apoptosis in Crohn's disease. *Inflamm Bowel Dis* 2006; 12: 346-54.
58. Watkins IR, Milligan ED, Maier SF. Glial activation: a driving force for pathological pain. *Trends Neurosci* 2001; 24: 450-55.
59. Rossi J, Herzig KH, Voikar V, Hiltunen PH, Segerstrale M, Airaksinen MS. Alimentary tract innervation deficits and dysfunction in mice lacking GDNF family receptor alfa2. *J Clin Invest* 2003; 112: 707-16.
60. Nasser Y, Fernandez E, Keenan CM, Ho W, Oland LD, Tibbles LA, et al. The role of enteric glia in intestinal physiology: the effects of the gliotoxin fluorocitrate on motor and secretory function. *Am J Physiol* 2006; 291: G912-27.
61. Burnstock G, Lavin S. Interstitial cells of Cajal and purinergic signalling. *Auton Neurosci* 2002; 97: 68-72.
62. Bassotti G, Villanacci V, Nascimbeni R, Cadei M, Manenti S, Sabatino G, et al. Colonic mast cells in controls and slow transit constipation patients. *Aliment Pharmacol Ther* 2011; 34: 92-99.
63. Bassotti G, Villanacci V, Fisogni S, Cadei M, Galletti A, Morelli A, et al. Comparison of three methods to assess enteric neuronal apoptosis in patients with slow transit constipation. *Apoptosis* 2007; 12: 329-32.
64. Bassotti G, Villanacci V, Fisogni S, Cadei M, Di Fabio F, Salerni B. Apoptotic phenomena are not a major cause of enteric neuronal loss in constipated patients with dementia. *Neuropathology* 2007; 27: 67-72.
65. Sechi LA, Ruehl A, Ahmed N, Usai D, Paccagnini D, Felis GE, et al. *Mycobacterium avium* subspecies paratuberculosis infects and multiplies in enteric glial cells. *World J Gastroenterol* 2007; 13: 5731-35.
66. Marruchella G, Ligios C, Albanese V, Cancedda MG, Madau L, Lalatta-Costerbosa G, et al. Enteroglia and neuronal involvement without apparent neuron loss in ileal enteric nervous system plexuses from scrapie-affected sheep. *J Gen Virol* 2007; 88: 2899-904.

67. Albanese V, Lawson VA, Hill AF, Cappai R, Di Guardo G, Staikopoulos V, et al. Evidence for prion protein expression in enteroglia cells of the myenteric plexus of mouse intestine. *Auton Neurosci* 2008; 140: 17-23.
68. di Giancamillo A, Vitari F, Bosi G, Savoini G, Domeneghini C. The chemical code of porcine enteric neurons and the number of enteric glial cells are altered by dietary probiotics. *Neurogastroenterol Motil* 2010; 22: e271-78.
69. Phillips RJ, Powley TL. Innervation of the gastrointestinal tract: patterns of aging. *Auton Neurosci* 2007; 136:1-19.
70. Wiskur B, Greenwood-Van Meerveld B. The aging colon: the role of enteric neurodegeneration in constipation. *Curr Gastroenterol Rep* 2010; 12: 507-12.
71. Bernard CE, Gibbons SJ, Gomez-Pinilla PJ, Lurken MS, Schmalz PF, Roeder JL, et al. Effect of age on the enteric nervous system of the human colon. *Neurogastroenterol Motil* 2009; 21: 746-e46.
72. Villanacci V, Bassotti G, Cathomas G, Maurer CA, Di Fabio F, Bisogni S, et al. Is pseudomelanosis coli a marker of colonic neuropathy in severely constipated patients? *Histopathology* 2006; 49: 132-37.
73. Rossi E, Villanacci V, Fisogni S, Morelli A, Salerni B, Grigolato P, et al. Chromosomal study of enteric glial cells and neurons by fluorescence in situ hybridization in slow transit constipation. *Neurogastroenterol Motil* 2007; 19: 578-84.
74. Bassotti G, Villanacci V. The London Classification of gastrointestinal neuromuscular pathology: a little more flexibility would be wise... *Gut* 2010 Sep 20.