



The Effect of Chlorhexidine on the Push-Out Bond Strength of Calcium-Enriched Mixture Cement

Fereshte Sobhnamayan^a, Alireza Adl^{a*}, Nooshin Sadat Shojaee^a, Samina Gavahian^b

^a Department of Endodontics, Dental School, Shiraz University of Medical Sciences, Shiraz, Iran; ^b Students' Research Committee, Dental School, International Branch, Shiraz University of Medical Sciences, Shiraz, Iran

ARTICLE INFO

Article Type:

Original Article

Received: 11 Aug 2014

Revised: 19 Nov 2014

Accepted: 03Dec 2014

*Corresponding author:
Alireza Adl, Department of
Endodontics, Dental School,
Shiraz University of Medical
Sciences, Shiraz, Iran

Tel: +98-711 6263193

Fax: + 98-7116270325

E-mail: adla@sums.ac.ir

ABSTRACT

Introduction: The aim of this *in vitro* study was to evaluate the effect of 2% chlorhexidine (CHX) on the push-out bond strength (BS) of calcium-enriched mixture (CEM) cement. **Methods and Materials:** Root-dentin slices from 60 single-rooted human teeth with the lumen diameter of 1.3 mm were used. The samples were randomly divided into 4 groups ($n=15$), and their lumens were filled with CEM cement mixed with either its specific provided liquid (groups 1 and 3) or 2% CHX (groups 2 and 4). The specimens were incubated at 37°C for 3 days (groups 1 and 2) and 21 days (groups 3 and 4). The push-out BS were measured using a universal testing machine. The slices were examined under a light microscope at 40× magnification to determine the nature of bond failure. The data were analyzed using the two-way ANOVA. For subgroup analysis the student t-test was applied. The level of significance was set at 0.05. **Results:** After three days, there was no significant difference between groups 1 and 2 ($P=0.892$). In the 21-day specimens the BS in group 3 (CEM) was significantly greater than group 4 (CEM+CHX) ($P=0.009$). There was no significant difference in BS between 3 and 21-day samples in groups 2 and 4 (CEM+CHX) ($P=0.44$). However, the mean BS after 21 days was significantly greater compared to 3-day samples in groups 1 and 3 ($P=0.015$). The bond failure in all groups was predominantly of cohesive type. **Conclusion:** Mixing of CEM with 2% CHX had an adverse effect the bond strength of this cement.

Keywords: Bond Strength; Calcium-Enriched Mixture; CEM Cement; Chlorhexidine; Push-out Bond strength; Root-End Filling Materials

Introduction

Calcium-enriched mixture cement (CEM) is a favorable biomaterial for repairing root perforations because of its excellent biocompatibility, sealing ability, hard tissue induction, cementogenesis and PDL formation [1-4]. This cement has antibacterial effect similar to calcium hydroxide [5]. It also has low cytotoxicity similar to mineral trioxide aggregate (MTA) [6]. CEM and tooth-colored ProRoot MTA showed similar sealing ability in repair of furcal perforation [7] or filling the entire canal space prior to root-end resection [8]. CEM cement has an acceptable fungicidal effects against *Candida Albicans* and is able to maintain its effect in concentration of 50 mg/mL after 24 h [9]. Different treatment strategies were applied for sealing perforation with root-end filling materials [10, 11].

Depending on circumstances, perforation site can be sealed after or prior to root canal cleaning and shaping [12]. However,

since leakage of some irrigants through the perforated area may cause severe irritation of the periodontal tissue during the cleaning and shaping of root canals [13], it has been suggested that the perforation defects should be repaired before complementing endodontic treatment [14]. Following the repair of furcal perforations, and after 7 days of incubation for initial set, endodontic treatment is performed with various irrigation solutions that cause inevitable contact of endodontic irrigants with the site of furcal repair [15].

Chlorhexidine (CHX) is a dicationic bisguanide chlorophenyl ring, that was initially used as a general disinfectant because of its broad antibacterial action [16]. In endodontics, CHX is also used as an irrigant to disinfect the root canal system [17, 18]. It has been shown that mixing MTA with CHX increases the antibacterial efficacy of MTA [19, 20]. A study in rats showed that MTA mixed with CHX caused only a weak inflammatory response on subcutaneous connective tissues, which subsided continuously over time; therefore, the

set mixture is considered biocompatible [21]. There are inconsistent results in the literature regarding effect of CHX on the physical properties of MTA. One study reported that MTA mixed with CHX gel did not set even after seven days [22]. In contrast, in the study by Holt *et al.* [19], MTA was mixed with liquid 2% CHX, and after 72 h most samples were set enough to allow performing compressive strength test. Their results revealed that MTA mixed with sterile water had compressive strength higher than that of MTA mixed with CHX. In an *in vitro* study, immersion of dentin disks filled with MTA in CHX had no significant effect on bond strength (BS) of MTA [15].

Different studies have been shown that CEM cement has higher antibacterial properties compared to MTA [5, 23]. Similar to MTA, it has been reported that mixing with CHX increases the antimicrobial effects of CEM cement [24].

The question is whether CHX affects the physical properties of CEM cement, or not. Resistance of dental materials to dislodgment forces is an important factor in the success of different endodontic procedures like repair of perforations, apical barrier formation, and root-end filling. Evaluation of the BS between these materials and dentin will show the value of adhesion between them. Different techniques can be used to evaluate the BS of a dental material to dentin including tensile, shear, and push-out BS tests. In the present study, push-out BS test was used, which is the most reliable method for evaluating the resistance of materials to dislodgment forces based on the results of previous studies [25, 26]. Therefore this study was designed to evaluate the effect of CHX on the push-out BS of CEM cement.

Materials and Methods

Freshly extracted, single-rooted human teeth, including maxillary incisors and mandibular premolars, were selected and stored in 0.5% chloramine-T before use. All the teeth had mature apices and intact roots. Teeth with cracks or internal resorption were excluded from the study. The crowns of all teeth were removed, and the middle thirds of the roots were sectioned perpendicular to the long axis to obtain 60 dentin disks with a thickness of 1.3 ± 0.2 mm.

The lumens of the dentin disks were prepared with sizes #2 to 5 of Gates Glidden drills (Dentsply Maillefer, Ballaigues, Switzerland), to obtain a standardized diameter of 1.3 mm. To remove the smear layer, disks were immersed in 17% ethylenediaminetetraacetic acid (EDTA), and then in 2.5% sodium hypochlorite (NaOCl), for 3 min each. The samples were then immediately washed in distilled water and dried. The dentin disks were randomly divided into 4 groups ($n=15$), and their lumens were filled with CEM cement (BioniqueDent, Tehran, Iran). In groups 1 and 3 (CEM), CEM cement was mixed according to the manufacturer's instruction; in groups 2 and 4 (CEM+CHX), CEM was mixed with 2% CHX (Consepsis V, Ultradent Products, Inc., South Jordan, UT, USA).

The specimens were wrapped in pieces of gauze soaked in normal saline and kept in sealed plastic containers. The specimens were incubated at 37°C for 3 (groups 1 and 2) and 21 days (groups 3 and 4).

Push-out bond strength test

After the experimental periods, the push-out BSs were measured using universal testing machine (Z050, Zwick GmbH, Ulm, Germany). The dentin disks were placed on a metal slab with a central hole to allow for the free motion of the plunger.

The specimens were loaded with a 0.7-mm diameter cylindrical stainless steel plunger at a speed of 1 mm/min [25]. The maximum load applied to CEM cement before dislodgement, was recorded in Newton (N). To express the BS in MPa, the recorded values in N was divided by the adhesion surface area of CEM cement in mm² calculated according to following formula; $2\pi r \times h$, where π is the constant 3.14, r is the root canal radius (1.3 mm), and h is the thickness of the root slice in mm.

The slices were then examined by the light microscope at 40× magnification to determine the nature of the bond failure. Each sample was categorized into 1 of 3 failure modes; adhesive failure at the CEM and dentin interface, cohesive failure within CEM cement and mixed failure.

Push-out BS data was transformed using natural logarithm to achieve normality. The two-way ANOVA test was used to assess the simultaneous effects of group and time. For subgroup analysis, the student's t-test was used. The level of significance was set at 0.05.

Results

Logarithm transformation was done to normalize data. The mean values and standard deviation of push-out BS in four experimental groups are shown in Table 1.

There was an interaction effect between all groups ($P=0.028$). Subgroup analysis showed that after 3 days, there was no significant difference between groups 1 (CEM) and 2 (CEM+CHX) ($P=0.892$). However, after 21 days, the BS in group 3 (CEM), was significantly more than group 4 (CEM+CHX) (0.92 ± 0.68 and 0.007 ± 1.06 , respectively) ($P=0.009$).

Moreover, there was not a significant difference in BS between groups 2 and 4 (CEM+CHX) ($P=0.44$). Comparison of groups 1 and 3 (CEM) showed that the mean BS after 21 days (0.92 ± 0.68) was significantly greater than that of 3-day samples (0.21 ± 0.81) ($P=0.015$).

Table 1. Mean (SD) of bond strength in different groups

Group (days)	Mean (SD)	SD
CEM+CHX (3)	1.55 (0.25)	0.78 (0.73)
CEM+CHX (21)	1.63 (0.007)	1.73 (1.05)
CEM (3)	1.70 (0.21)	1.52 (0.80)
CEM (21)	3.03 (0.92)	1.73 (0.68)

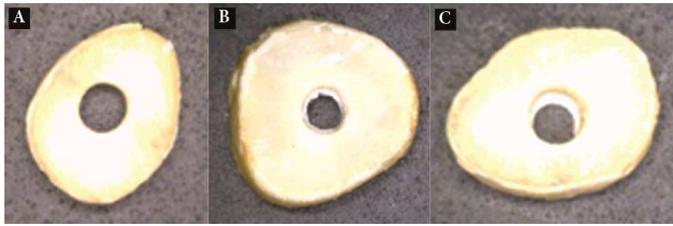


Figure 1. Different failure modes; A) adhesive failure at the CEM and dentin interface, B) cohesive failure within CEM cement, C) mixed failure

Bond failure in all groups was predominantly of cohesive type, although some samples exhibited mixed and adhesive failure patterns, as well (Table 2) (Figure 1).

Discussion

This *in vitro* study compared the push-out BS of CEM cement samples mixed with 2% CHX to that of conventionally mixed samples, at two different time intervals and showed that mixing CEM cement with CHX reduces its BS.

The success of furcal perforation repair depends on a well-placed coronal restoration and the resistance of the repair material to displacement forces during condensation of permanent restorative materials. The amalgam condensation force could reach up to 3.7-11.3 MPa during condensation with different pluggers [27]. This pressure is high enough to cause the dislodgement of furcal repair materials [25, 28]. Thus, the BS of the perforation repair materials is an important factor in clinical situations. To assess the BS, various methods have been used including tensile, shear, and push-out BS tests [29]. The push-out bond test has been shown to be practical and reliable [30-33].

The results of the present study showed that there was no statistically significant difference between the push-out BS of groups 1 and 2 (*i.e.* CEM and CEM+CHX) at 3-day interval. This finding is in accordance with those of Yan *et al.* [15] who found that immersion of MTA in CHX for two hours, did not adversely affect MTA-dentin BS. Holt *et al.* [19] also showed that mixing of MTA with 2% CHX had no adverse effect on the compressive strength of MTA after three days.

However, after 21 days the results showed that the BS in group 3 (CEM) was significantly greater than group 4 (CEM+CHX). Increasing the setting time from 3 to 21 days, did not increase the push-out BS of CEM+CHX mixture contrary to that of CEM.

Evaluation of the effect of time on the BS of MTA in some studies, has shown that in dry conditions there is no significant increase in the BS of MTA but under wet conditions the push-out BS of MTA showed a statistically significant increase from days 3 to 21 [34]. The results of the current study is also partly in agreement with the results of Rahimi *et al.* [31], who reported an increase in the BS of CEM cement mixed with normal saline from 24 h to 7 days although in the current study CEM+CHX group showed an increase in the BS with the pass of time. One reason for the observed discrepancies in groups 1 and 2 in the present study may be related to the probable morphological

alteration of the interfacial layer caused by CHX, as it has been reported that CHX caused a significant increase in the amount of the needle-shaped crystals in MTA [15].

Hong *et al.* [35] found that when CHX was used as an irrigant, apparent crystalline structures on the surface of both accelerated and nonaccelerated MTA samples were not observed. The surface crystals had the thin plate structures, and their size was reduced almost to one tenth of those of the control group. In an energy dispersive spectroscopy (EDS) analysis, silicon was detected along with calcium, oxygen, and carbon, which proved that they were not the typical CH crystals. These findings may explain why the push-out BS of the CHX groups was significantly reduced in MTA samples exposed to CHX. This could also be a reason for the lower BS of CHX group in this study; however, scanning electron microscopy (SEM) analysis of CEM cement after exposure to CHX is recommended.

In the present study, the bond failure in all groups was predominantly of cohesive type, although some samples exhibited mixed and adhesive failure patterns (Table 2). This result is in accordance with those of Guner *et al.* [36] who showed that Dyract AP [a new hydrophilic glass-ionomer cement (DeTrey/Dentsply, Konstanz, Germany)], and Biodentin (Septodont, Saint-Maur-des-Fosses, France) showed predominantly cohesive bond failure of when exposed to different irrigating solutions such as CHX and normal saline. But for MTA the failure pattern was mostly of the adhesive type [36] which is not in accordance with the present study. These differences in the pattern of bond failures might be attributed to different dental materials used in different studies and their different chemical composition and different particle sizes. As CEM cement consists of higher percentage of small particle sizes [37]. Smaller particle size of root-end filling materials like nano MTA showed better physical and chemical properties. It also shows increased surface area and less porosity when exposed to acidic pH levels [38]. The compressive strength of nano MTA is also less affected by acidic environment [39]. The faster setting time of the CEM cement may cause a shorter working time for this material and a faster chemical reaction which is the most important period for structure formation and ion release [40].

It has also been shown that CHX can be adsorbed onto hydroxyapatite and teeth (substantivity property of CHX) [31] and it may improve the BS of MTA to dentinal walls [31]. This also could happen in CEM cement samples and cause the bond failure in groups to be of cohesive type. However SEM analysis of CEM cement samples is needed to prove these theories.

Table 2. Type of bond failure in different groups

Group (days)	Adhesive (%)	Cohesive (%)	Mixed (%)
CEM+CHX (3)	13.33	86.66	0
CEM+CHX (21)	6.66	93.33	0
CEM (3)	6.66	66.66	26.66
CEM (21)	0	100	0

Conclusion

Mixing CEM cement with 2% CHX had an adverse effect on its bond strength. It is therefore not considered a suitable substitute for CEM liquid in clinical situations where the cement may be subjected to dislodgement forces.

Acknowledgment

This article is based on a thesis conducted by Dr. S. Gavahian. The authors thank the Research Vice Chancellor of Tabriz University of Medical Sciences for supporting this research. The authors are also grateful to Dr. M. Vossoghi from the Dental Research Development Center for the statistical analysis.

Conflict of Interest: 'None declared'.

References

1. Asgary S, Eghbal MJ, Parirokh M, Ghanavati F, Rahimi H. A comparative study of histologic response to different pulp capping materials and a novel endodontic cement. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod.* 2008;106(4):609-14.
2. Asgary S, Eghbal MJ, Parirokh M. Sealing ability of a novel endodontic cement as a root-end filling material. *J Biomed Mater Res A.* 2008;87(3):706-9.
3. Asgary S, Eghbal MJ, Ehsani S. Periradicular regeneration after endodontic surgery with calcium-enriched mixture cement in dogs. *J Endod.* 2010;36(5):837-41.
4. Tabarsi B, Parirokh M, Eghbal MJ, Haghdoost AA, Torabzadeh H, Asgary S. A comparative study of dental pulp response to several pulpotomy agents. *Int Endod J.* 2010;43(7):565-71.
5. Asgary S, Kamrani FA. Antibacterial effects of five different root canal sealing materials. *J Oral Sci.* 2008;50(4):469-74.
6. Mozayeni MA, Milani AS, Marvasti LA, Asgary S. Cytotoxicity of calcium enriched mixture cement compared with mineral trioxide aggregate and intermediate restorative material. *Aust Endod J.* 2012;38(2):70-5.
7. Haghgoo R, Niyakan M, Nazari Moghaddam K, Asgary S, Mostafaloo N. An In vitro Comparison of Furcal Perforation Repaired with Pro-root MTA and New Endodontic Cement in Primary Molar Teeth- A Microleakage Study. *J Dent (Shiraz).* 2014;15(1):28-32.
8. Milani AS, Shakouie S, Borna Z, Sighari Deljavan A, Asghari Jafarabadi M, Pournaghi Azar F. Evaluating the Effect of Resection on the Sealing Ability of MTA and CEM Cement. *Iran Endod J.* 2012;7(3):134-8.
9. Kangarlou A, Sofiabadi S, Yadegari Z, Asgary S. Antifungal effect of calcium enriched mixture cement against *Candida albicans*. *Iran Endod J.* 2009;4(3):101-5.
10. Main C, Mirzayan N, Shabahang S, Torabinejad M. Repair of root perforations using mineral trioxide aggregate: a long-term study. *J Endod.* 2004;30(2):80-3.
11. Menezes R, da Silva Neto UX, Carneiro E, Letra A, Bramante CM, Bernadinelli N. MTA repair of a supracrestal perforation: a case report. *J Endod.* 2005;31(3):212-4.
12. Torabinejad M, Chivian N. Clinical applications of mineral trioxide aggregate. *J Endod.* 1999;25(3):197-205.
13. Gernhardt CR, Eppendorf K, Kozłowski A, Brandt M. Toxicity of concentrated sodium hypochlorite used as an endodontic irrigant. *Int Endod J.* 2004;37(4):272-80.
14. CJ R. Nonsurgical endodontics retreatment. In: S C, RC B, editors. *Pathways of the pulp.* 8th ed. St. Louis: Mosby Inc; 2002. pp. 917.
15. Yan P, Peng B, Fan B, Fan M, Bian Z. The effects of sodium hypochlorite (5.25%), Chlorhexidine (2%), and Glyde File Prep on the bond strength of MTA-dentin. *Journal of endodontia.* 2006;32:58-60.
16. Hirst RC. Chlorhexidine: a review of the literature. *Periodontal Abstr.* 1972;20(2):52-8.
17. Karale R, Thakore A, Shetty V. An evaluation of antibacterial efficacy of 3% sodium hypochlorite, high-frequency alternating current and 2% chlorhexidine on *Enterococcus faecalis*: An in vitro study. *J Conserv Dent.* 2011;14(1):2-5.
18. Alves FR, Almeida BM, Neves MA, Moreno JO, Rôças IN, Siqueira JF. Disinfecting oval-shaped root canals: effectiveness of different supplementary approaches. *J Endod.* 2011;37(4):496-501.
19. Holt DM, Watts JD, Beeson TJ, Kirkpatrick TC, Rutledge RE. The anti-microbial effect against *enterococcus faecalis* and the compressive strength of two types of mineral trioxide aggregate mixed with sterile water or 2% chlorhexidine liquid. *J Endod.* 2007;33(7):844-7.
20. Stowe TJ, Sedgley CM, Stowe B, Fenno JC. The effects of chlorhexidine gluconate (0.12%) on the antimicrobial properties of tooth-colored ProRoot mineral trioxide aggregate. *J Endod.* 2004;30(6):429-31.
21. Sumer M, Muglali M, Bodrumlu E, Guvenc T. Reactions of connective tissue to amalgam, intermediate restorative material, mineral trioxide aggregate, and mineral trioxide aggregate mixed with chlorhexidine. *J Endod.* 2006;32(11):1094-6.
22. Kogan P, He J, Glickman GN, Watanabe I. The effects of various additives on setting properties of MTA. *J Endod.* 2006;32(6):569-72.
23. Asgary S, Akbari Kamrani F, Taheri S. Evaluation of antimicrobial effect of MTA, calcium hydroxide, and CEM cement. *Iran Endod J.* 2007;2(3):105-9.
24. Bidar M, Naderinasab M, Talati A, Ghazvini K, Asgari S, Hadizadeh B, Gharechahi M, Mashadi NA. The effects of different

- concentrations of chlorhexidine gluconate on the antimicrobial properties of mineral trioxide aggregate and calcium enrich mixture. *Dent Res J (Isfahan)*. 2012;9(4):466-71.
25. Saghiri MA, Shokouhinejad N, Lotfi M, Aminsobhani M, Saghiri AM. Push-out bond strength of mineral trioxide aggregate in the presence of alkaline pH. *J Endod*. 2010;36(11):1856-9.
 26. Shokouhinejad N, Nekoofar MH, Iravani A, Kharrazifard MJ, Dummer PM. Effect of acidic environment on the push-out bond strength of mineral trioxide aggregate. *J Endod*. 2010;36(5):871-4.
 27. Lussi A, Brunner M, Portmann P, Buerger W. Condensation pressure during amalgam placement in patients. *Eur J Oral Sci*. 1995;103(6):388-93.
 28. Hashem AA, Wanees Amin SA. The effect of acidity on dislodgment resistance of mineral trioxide aggregate and bioaggregate in furcation perforations: an in vitro comparative study. *J Endod*. 2012;38(2):245-9.
 29. Loxley EC, Liewehr FR, Buxton TB, McPherson JC. The effect of various intracanal oxidizing agents on the push-out strength of various perforation repair materials. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 2003;95(4):490-4.
 30. Goracci C, Tavares AU, Fabianelli A, Monticelli F, Raffaelli O, Cardoso PC, Tay F, Ferrari M. The adhesion between fiber posts and root canal walls: comparison between microtensile and push-out bond strength measurements. *Eur J Oral Sci*. 2004;112(4):353-61.
 31. Rahimi S, Ghasemi N, Shahi S, Lotfi M, Froughreyhani M, Milani AS, Bahari M. Effect of blood contamination on the retention characteristics of two endodontic biomaterials in simulated furcation perforations. *J Endod*. 2013;39(5):697-700.
 32. Shokouhinejad N, Hoseini A, Gorjestani H, Shamshiri AR. The effect of different irrigation protocols for smear layer removal on bond strength of a new bioceramic sealer. *Iran Endod J*. 2013;8(1):10-3.
 33. Salem Milani A, Froughreyhani M, Charchi Aghdam S, Pournaghiazar F, Asghari Jafarabadi M. Mixing with propylene glycol enhances the bond strength of mineral trioxide aggregate to dentin. *J Endod*. 2013;39(11):1452-5.
 34. Gancedo-Caravia L, Garcia-Barbero E. Influence of humidity and setting time on the push-out strength of mineral trioxide aggregate obturations. *J Endod*. 2006;32(9):894-6.
 35. Hong ST, Bae KS, Baek SH, Kum KY, Shon WJ, Lee W. Effects of root canal irrigants on the push-out strength and hydration behavior of accelerated mineral trioxide aggregate in its early setting phase. *J Endod*. 2010;36(12):1995-9.
 36. Guner MB, Akbulut MB, Eldeniz AU. Effect of various endodontic irrigants on the push-out bond strength of biodentine and conventional root perforation repair materials. *J Endod*. 2013;39(3):380-4.
 37. Soheilipour E, Kheirieh S, Madani M, Akbarzadeh Baghban A, Asgary S. Particle size of a new endodontic cement compared to Root MTA and calcium hydroxide. *Iran Endod J*. 2009;4(3):112-6.
 38. Saghiri MA, Asgar K, Lotfi M, Garcia-Godoy F. Nanomodification of mineral trioxide aggregate for enhanced physiochemical properties. *Int Endod J*. 2012;45(11):979-88.
 39. Saghiri MA, Garcia-Godoy F, Asatourian A, Lotfi M, Banava S, Khezri-Boukani K. Effect of pH on compressive strength of some modification of mineral trioxide aggregate. *Med Oral Patol Oral Cir Bucal*. 2013;18(4):e714-20.
 40. Asgary S, Shahabi S, Jafarzadeh T, Amini S, Kheirieh S. The properties of a new endodontic material. *J Endod*. 2008;34(8):990-3.

Please cite this paper as: Sobhnamayan F, Adl A, Sadat Shojaee N, Gavahian S. The Effect of Chlorhexidine on the Push-Out Bond Strength of Calcium-Enriched Mixture Cement. *Iran Endod J*. 2014;10(1): 59-63.