

Impact of proteomics investigations on gastric cancer treatment and diagnosis

Mohammad Rostami-Nejad¹, Mostafa Rezaei-Tavirani², Vahid Mansouri², Zahra Akbari³, Saeed Abdi¹

¹Gastroenterology and Liver Diseases Research Center, Research Institute for Gastroenterology and Liver Diseases, Shahid Beheshti University of Medical Sciences, Tehran, Iran

²Proteomics Research Center, Faculty of Paramedical Sciences, Shahid Beheshti University of Medical Sciences, Tehran, Iran

³Laser Application in Medical Sciences Research Center, Shahid Beheshti University of Medical Sciences, Tehran, Iran

ABSTRACT

Gastric cancer is one of the epidemic diseases with a high mortality rate in different countries. It causes many health problems in the world every year. It affects the digestive tract, and in advanced cases, its treatment has many difficulties. Early detection of cancer in different parts of the gastrointestinal tract can be accompanied by inexpensive treatment. As cancer cells make different biomarkers during different stages of the disease, researchers are looking for different biomarkers for gastrointestinal cancers detection. On the other hand, with the advent of advanced techniques such as proteomics and the discovery of a large number of proteins related to gastrointestinal cancer, finding the role of these proteins is essential. Indeed, the function of large amounts of these proteins has remained unknown.

Data from databases such as genes and proteins associated with gastrointestinal cancers were collected and the proteomic data of these databases were analyzed to find a clear perspective of the impact of proteomics in gastric cancer management.

The role of heat shock proteins, metabolic proteins, membrane binding proteins, galectins, prohibitins, S100 proteins, and many different types of proteins in gastric cancer was highlighted. This article reviewed proteomic researches in cancer-related areas of the gastric cancer in order to evaluate the findings of researchers.

Keywords: Gastric cancer, Biomarker, Proteomic.

(Please cite as: Rostami-Nejad M, Rezaei-Tavirani M, Mansouri V, Akbari Z, Abdi S. Impact of proteomics investigations on gastric cancer treatment and diagnosis. *Gastroenterol Hepatol Bed Bench* 2019;12(Suppl. 1):S1-S7).

Introduction

Gastric cancer mortality is the second prevalent cancer in the world (1, 2). Early onset gastric cancer pathogenesis is still unclear (2). Most gastric cancers occur in a sporadic manner and over the age of 45 years old in affected people (3). Approximately 5% of patients survive longer than 5 years after treatments in the patients diagnosed in the late stages of GC. Thus, there is motivation for early GC detection of non-invasive biomarkers, earlier than development of GC metastasis (4). Extensive research has been done on

tissue (5), blood (6), and body fluids as well as tumor cells (7) to find gastric cancer (GC) biomarkers (6), with emphasis on finding proteins, DNA (8), and RNA (9). Early detection of GC could assist in suitable treatment of disease (10). Recently, proteomics technology as an efficient tool has been able to help researchers to identify different proteins involved in GC (11).

There are not any statistical evidence to identify GC biomarkers and proteomics researches could assist in identifying the molecular basis of GC (12). Understanding the mechanism of GC is one of the goals which proteomics could contribute to (11). One of the methods used in the proteomics is mass spectrometry (MS), and the researchers have come up with several findings via this technique on GC. Yang J et al. used

Received: 14 October 2019 Accepted: 8 December 2019

Reprint or Correspondence: Mostafa Rezaei-Tavirani, PhD. *Proteomics Research Center, Faculty of Paramedical Sciences, Shahid Beheshti University of Medical Sciences, Tehran, Iran.*

E-mail: tavirany@yahoo.com

ORCID ID: 0000-0003-1767-7475

S2 Impact of proteomics investigations on gastric cancer treatment and diagnosis

magnetic based purification and matrix-assisted laser desorption/ionization time-of-flight (MALDI-TOF) mass spectrometry in the serum of GC patients to identify SERPINA1 and ENOSF as potential GC protein biomarkers (13). Different sources and techniques were used to identify biomarkers of GC. WU W et al. represented a simple MS-based scoring for biomarkers to detect the early period of GC from gastric fluid and serum (4). They found relations between the levels of PRB2, SERPINA1, Elastase 3A, CystD, and CELA3B expression in GC progression (4). They believed that three-biomarker panel of CystD+PepA-Ela3A could be sufficient for GC diagnosis with 95.7% sensitivity. The following parts refer to the highlighted findings about the role of different types of proteins in GC onset and development via proteomics.

Methods

Data were obtained from PubMed, Scopus, and Google scholar, from 105 articles. The search process is shown in Figure 1. Among the 105 full texts, 65 documents were selected to review.

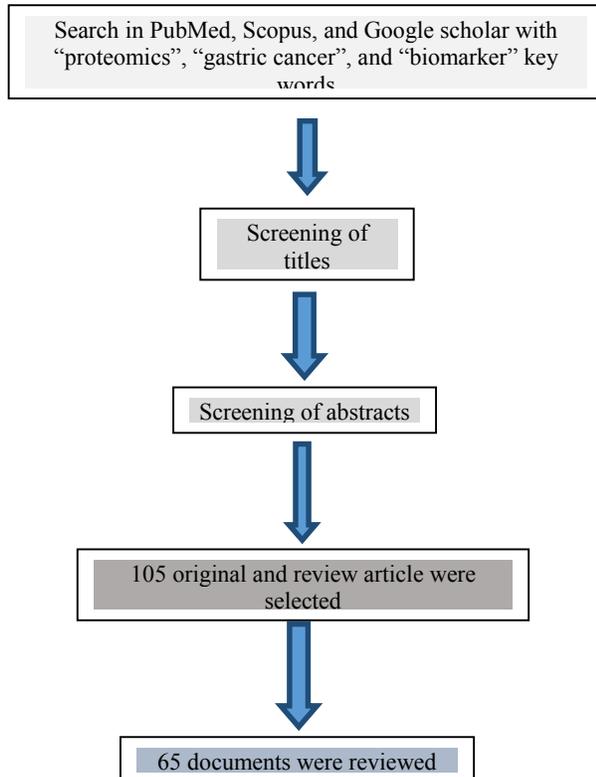


Figure 1. Schematic representation of the search process

Results

Heat Shock Proteins

There are some heat shock proteins reported in several proteomics approaches as heat shock protein 27 (HSP 27) with overexpression in GC (14-17). Other studies revealed the upregulation of other heat shock proteins such as HSB60 (18) and HSB70 (19). In one study, the overexpression of HSP70 was observed in GC treated by administration of N-methyl-N'-nitro-N-nitrosoguanidine in rats (20). Other researchers have reported the role of HSP90 (21) and HSP105 (21, 22) in GC. HSPs are chaperons cells synthesize against stressful stimuli for survival of cells (23). However, there has been no evidence of a link between HSO70 expression and pathological characteristics (12). For example, in one study, diminished expression of HSP27 in GC cells derived from lymph nodes compared to GC cells from tumors was reported (24), while the relationship between HSP27 and the size of tumor as well as distant of metastasis was mentioned by other articles (25). The increased expression of HSP60 and HSP90 has been reported in invasive cell line compared to non-invasive cell line (21). On the other hand, decreased expression of HSP90 in GC cells derived from lymph node metastasis is inconsistent with other studies in increasing its expression (24). The drug inhibitors of HSP90 were tested clinically and it seems they could affect GC cells (26). Research has suggested that the combination of HSP inhibitors with other anticancer drugs could be beneficial in clinics to treat GC (26).

Metabolic proteins

Proteomic studies have revealed the overexpression of ENOA protein in GC cells compared to nonneoplastic gastric cells with or without lymph node metastasis (16,28). Capello M et al. suggested the role and function of ENOA in the metastasis process of GC and its functions (28). The diminished expression of GKN1 was reported in GC cells and ENOA protein could regulate GKN1 activity (17,29). Through the downregulation of ENOA, GC cell cycle was arrested and it was equal to overexpression of GKN1 (30). ENOA overexpression leads to tumor growth by glycolysis and pyruvate synthesis (28,31). Metabolic proteins of Krebs cycle and oxidative phosphorylation were downregulated in GC cells, according to

proteomic studies (32, 33). The metabolic profile of GC cells is different to non-metastatic gastric cell profiles (15). Proteomic studies suggested Warburg effect in GC cells by forming lactate from glucose through glycolysis (34). Glucose oxidation is essential for synthesis of proteins and lipids as well as nucleic acids during cell divisions where high glycolysis is an advantage for GC tumor cells (35, 36). Forced transition of the Krebs cycle from glycolysis process could be used as a treatment in gastric cancer (15).

Membrane binding proteins

Anexins, as membrane binding proteins, are calcium dependent. These intracellular proteins can form membrane bond plexus in the surface of cell membrane to interact with other proteins for different membrane functions as differentiation, migration, and dynamics of membrane (37). The increase in ANXA2 has been reported by several proteomics analyses in GC cells (32,38). ANXA2 expression increases in tumors with lymph node metastasis as compared to non-neoplastic gastric cancer cells (16). Another study revealed ANXA2 overexpression in invasive GC cells compared to non-invasive cells (39). Tumor size and location, differentiation, vessel invasion, and lymph node metastasis could affect ANXA2 overexpression (40). Its overexpression could maintain the malignancy and motility of GC cells (41). Some proteomic studies reported diminished expression of ANXA1 in GC cells (21), but other studies in contrast reported overexpression of ANXA1 (16). Nevertheless, ANXA1 overexpression leads to GS invasion as well as lymph node metastasis, and it is linked with prognostic factors such as venous and lymphatic invasions and advanced stages of GC (42, 43). However, other studies suggest ANXA1 expression during the early stages of gastric cancer (44). Proteomics investigation results have demonstrated overexpression of ANXA3, ANXA5, and ANXA13 in GC cells (16,45-47). Decreased expression of ANXA3 would suppress migration and invasion characteristics of GC cells (48). ANXA6, as a tumor suppressor factor of GC cells, acts through promotor mutilation (498). The overexpression of ANAXA7 in GC patients leads to reduction of survival rate vice versa (50). Reduction of ANAXA10 in GC cells and low survival rate have been reported previously. A proteomic analysis revealed the

regulatory duty of ANAXA10 in GC cells proliferation (551). It is suggested that ANAXA 10 may act as a tumor suppressor in GC cells, and its expression as ANAXA7 in intestinal and diffused type of GC cells is not similar (52). These results remarkably demonstrate the prominent action of ANAXA s in GC cells development.

Galectins roles

Galectins (GLA) could have a role in GC development by resistance to cell death, continuing proliferative signaling and resistance to cell death as well as activation of metastasis (53). GLA1 expression in metastatic cell line is associated with the size of tumor and metastasis of lymph node as well as survival rate of GC patients (54, 55). Proteomic approaches have revealed GLA4 and GLA2 overexpression in GC cells (32, 56). GLA2 overexpression is associated with advanced stages of GC and lymph node metastasis; thus, loss of GAL2 could play an important role in GC aggression (56). GLA3 expression in GC is reduced which is associated with distant metastasis (39). Poor expression of GLA3 equals to poorer prognosis of GC and other types of cancer (57-59). Several studies are required to improve the treatment of GC with expression changes of galectins.

S100 proteins

S100 proteins are involved in several biological functions in cells/as proliferation and motility (60) plus chemotactic and angiogenesis activities. Several S100 proteins could bind to annexins to form a cytoskeleton and are perhaps involved in cancer cell development (61). Overexpression of S100A2 protein could reduce the ability of GC cells to invade, and upon of reduction of S100a expression, the invasive ability of GC cells increased. Down-regulation of S100A2 protein in gastric carcinoma relative to the adjacent non-cancerous gastric tissues is reported by Ying Fu liu etal. (62)

Prohibitions

Proteomic approaches have revealed the overexpression of prohibitions in GC (63). However, other studies of GC showed decreased expression of prohibitions (64). Initiation of GC and tumor differentiation is a result of reduced expression of prohibitions (65).

S4 Impact of proteomics investigations on gastric cancer treatment and diagnosis

Table 1. The discussed proteins which were involved in gastric cancer are presented

R	Protein	Regulation	Ref.	Additional explanation
1	HSP27	up	14-17	It is down-regulated in GC cells derived from lymph nodes compare to GC cells from tumors (23).
2	HSP60	up	18, 21	
3	HSP70	up	19-20	
4	HSP90	up	21	
5	HSP105	up	22	It is down-regulated in GC cells derived from lymph node metastasis (23).
6	ENOA	up	16, 28	
7	GKN1	down	28	
8	ANXA1	Up and down	16	
9	ANXA2	up	32, 38	
10	ANXA3	up	16, 45-47	
11	ANXA5	up	16, 45-47	
12	ANXA6	down	49	
13	ANXA7	up	50	
14	ANXA10	down	51	
15	ANXA13	up	16, 45-47	
16	GLA1	up	54	
17	GLA2	down	55	
18	GLA3	down	39	
19	GLA4	up	32	
20	S100A2	down	62	
21	Prohibitins	up	63	Down regulation also is reported (62)
22	EPHA2	up	68	
23	CALD	down	24	
24	CAPG	up	69	
25	CRIP1	up	67	

Different types of proteins

Proteomic studies have identified other proteins involved in GC. Some of them are named as EPH receptor A2 (EPHA2), caldesmon (CALD), intestinal cysteine-rich protein 1 (CRIP1), and macrophage capping protein (CAPG)(66-68). Proteomic analysis has revealed CAPG overexpression in GC cells with lymph node metastasis (69). CALD expression was reduced in GC cells causing development of migration and invasion; thus it may have a critical role in progress of GC (24). The summary of findings is tabulated in the table 1.

Conclusion

The proteins emphatically highlighted in this review were found by high throughput screening methods. They could have a major role in GC. Proteomics technique may assist in understanding the mechanisms involved in tumor phenotype. Also, increasing the gastric carcinogenesis knowledge could assist in

improving the treatment methods. However, proteomics studies of GC in the elementary stages and long distance are still required to obtain the exact biomarkers involved in GC for essential useful diagnosis and treatment of disease. Meanwhile, the heterogeneity of tumors requires different several biomarkers where proteomics could assist in finding them.

Conflict of interests

The authors declare that they have no conflict of interest.

References

1. Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. *A Cancer J Clin* 2015;65:87-108.
2. Sitarz R, Skierucha M, Mielko J, Offerhaus GJA, Maciejewski R, Polkowski WP. Gastric cancer: epidemiology, prevention, classification, and treatment. *Cancer Manage Res* 2018;10:239.

3. Skierucha M, Milne AN, Offerhaus GJA, Polkowski WP, Maciejewski R, Sitarz R. Molecular alterations in gastric cancer with special reference to the early-onset subtype. *World Gastroenterol* 2016;22:2460.
4. Wu W, Yong WW, Chung MC. A simple biomarker scoring matrix for early gastric cancer detection. *Proteomics* 2016;16:2921-30.
5. Pan S, Brentnall TA, Kelly K, Chen R. Tissue proteomics in pancreatic cancer study: discovery, emerging technologies, and challenges. *Proteomics* 2013;13:710-21.
6. Werner S, Chen H, Tao S, Brenner H. Systematic review: serum autoantibodies in the early detection of gastric cancer. *Int J Cancer* 2015;136:2243-52.
7. Catenacci DV, Liao WL, Zhao L, Whitcomb E, Henderson L, O'Day E, et al. Mass-spectrometry-based quantitation of Her2 in gastroesophageal tumor tissue: comparison to IHC and FISH. *Gastric Cancer* 2016;19:1066-79.
8. Toiyama Y, Okugawa Y, Goel A. DNA methylation and microRNA biomarkers for noninvasive detection of gastric and colorectal cancer. *Biochem Biophys Res Commun* 2014;455:43-57.
9. Liu R, Zhang C, Hu Z, Li G, Wang C, Yang C, et al. A five-microRNA signature identified from genome-wide serum microRNA expression profiling serves as a fingerprint for gastric cancer diagnosis. *Eur J Cancer* 2011;47:784-91.
10. Xiao H, Zhang Y, Kim Y, Kim S, Kim JJ, Kim KM, et al. Differential Proteomic Analysis of Human Saliva using Tandem Mass Tags Quantification for Gastric Cancer Detection. *Sci Rep* 2016;6:22165.
11. Hu W, Wang J, Luo G, Luo B, Wu C, Wang W, et al. Proteomics-based analysis of differentially expressed proteins in the CXCR1-knockdown gastric carcinoma MKN45 cell line and its parental cell. *Acta Biochim Biophys Sin* 2013;45:857-66.
12. Leal MF, Wisniewski F, de Oliveira Gigeck C, Do Santos LC, Calcagno DQ, Burbano RR, et al. What gastric cancer proteomic studies show about gastric carcinogenesis? *Tumor Biol* 2016;37:9991-10010.
13. Yang J, Xiong X, Wang X, Guo B, He K, Huang C. Identification of peptide regions of SERPINA1 and ENOSF1 and their protein expression as potential serum biomarkers for gastric cancer. *Tumour Biol* 2015;36:5109-18.
14. Ryu JW, Kim HJ, Lee YS, Myong NH, Hwang CH, Lee GS, et al. The proteomics approach to find biomarkers in gastric cancer. *J Korean Med Sci* 2003;18:505.
15. Cai Z, Zhao JS, Li JJ, Peng DN, Wang XY, Chen TL, et al. A combined proteomics and metabolomics profiling of gastric cardia cancer reveals characteristic dysregulations in glucose metabolism. *Mol Cell Proteomics* 2010;9:2617-28.
16. Leal MF, Chung J, Calcagno DQ, Assumpcao PP, Demachki S, da Silva IDC, et al. Differential proteomic analysis of noncardia gastric cancer from individuals of northern Brazil. *PLoS One* 2012;7:e42255.
17. Kočevar N, Odreman F, Vindigni A, Grazio SF, Komel R. Proteomic analysis of gastric cancer and immunoblot validation of potential biomarkers. *World J Gastroenterol* 2012;18:1216.
18. Wu C, Luo Z, Chen X, Wu C, Yao D, Zhao P, et al. Two-dimensional differential in-gel electrophoresis for identification of gastric cancer-specific protein markers. *Oncol Rep* 2009;21:1429-37.
19. Cheng Y, Zhang J, Li Y, Wang Y, Gong J. Proteome analysis of human gastric cardia adenocarcinoma by laser capture microdissection. *BMC Cancer* 2007;7:191.
20. Chen J, Kähne T, Röcken C, Götz T, Yu J, Sung JJ, et al. Proteome analysis of gastric cancer metastasis by two-dimensional gel electrophoresis and matrix assisted laser desorption/ionization-mass spectrometry for identification of metastasis-related proteins. *J Proteome Res* 2004;3:1009-16.
21. Chen YR, Juan HF, Huang HC, Huang HH, Lee YJ, Liao MY, et al. Quantitative proteomic and genomic profiling reveals metastasis-related protein expression patterns in gastric cancer cells. *J Proteome Res* 2006;5:2727-42.
22. Nakatsura T, Senju S, Yamada K, Jotsuka T, Ogawa M, Nishimura Y. Gene cloning of immunogenic antigens overexpressed in pancreatic cancer. *Biochemical and biophysical research communications*. 2001;281(4):936-44.
23. Lianos GD, Alexiou GA, Mangano A, Mangano A, Rausei S, Boni L, et al. The role of heat shock proteins in cancer. *Cancer Lett* 2015;360:114-8.
24. Hou Q, Tan HT, Lim KH, Lim TK, Khoo A, Tan IB, et al. Identification and functional validation of caldesmon as a potential gastric cancer metastasis-associated protein. *J Proteome Res* 2013;12:980-90.
25. Kapranos N, Kominea A, Konstantinopoulos P, Savva S, Artelaris S, Vандoros G, et al. Expression of the 27-kDa heat shock protein (HSP27) in gastric carcinomas and adjacent normal, metaplastic, and dysplastic gastric mucosa, and its prognostic significance. *J Cancer Res Clin Oncol* 2002;128:426-32.
26. Liu H, Lu J, Hua Y, Zhang P, Liang Z, Ruan L, et al. Targeting heat-shock protein 90 with ganetespib for molecularly targeted therapy of gastric cancer. *Cell Death Dis* 2015;6:e1595.
27. Banerji U. O7. 6HSP90 inhibitors in the clinic. *Ann Oncol* 2015;26.
28. Capello M, Ferri-Borgogno S, Cappello P, Novelli F. α -enolase: a promising therapeutic and diagnostic tumor target. *FEBS J* 2011;278:1064-74.
29. Bai Z, Ye Y, Liang B, Xu F, Zhang H, Zhang Y, et al. Proteomics-based identification of a group of apoptosis-related proteins and biomarkers in gastric cancer. *Int J Oncol* 2011;38:375-83.
30. Yan GR, Xu SH, Tan ZL, Yin XF, He QY. Proteomics characterization of gastrokine 1-induced growth inhibition of gastric cancer cells. *Proteomics* 2011;11:3657-64.

S6 Impact of proteomics investigations on gastric cancer treatment and diagnosis

- 31.Pancholi V. Multifunctional α -enolase: its role in diseases. *Cell Mol Life Sci* 2001;58:902-20.
- 32.Kočevar N, Grazio SF, Komel R. Two-dimensional gel electrophoresis of gastric tissue in an alkaline pH range. *Proteomics* 2014;14:311-21.
- 33.Goh WQJ, Ow GS, Kuznetsov VA, Chong S, Lim YP. DLAT subunit of the pyruvate dehydrogenase complex is upregulated in gastric cancer-implications in cancer therapy. *Am J Trans Res* 2015;7:1140.
- 34.House SW, Warburg O, Burk D, Schade AL. On respiratory impairment in cancer cells. *Science* 1956;124:267-72.
- 35.Gatenby RA, Gillies RJ. Why do cancers have high aerobic glycolysis? *Nat Rev Cancer* 2004;4:891.
- 36.DeBerardinis RJ, Sayed N, Ditsworth D, Thompson CB. Brick by brick: metabolism and tumor cell growth. *Curr Opin Genet Dev* 2008;18:54-61.
- 37.Gerke V, Creutz CE, Moss SE. Annexins: linking Ca²⁺ signalling to membrane dynamics. *Nat Rev Mol Cell Biol* 2005;6:449.
- 38.Liu R, Li Z, Bai S, Zhang H, Tang M, Lei Y, et al. Mechanism of cancer cell adaptation to metabolic stress: proteomics identification of a novel thyroid hormone-mediated gastric carcinogenic signaling pathway. *Mol Cell Proteomics* 2009;8:70-85.
- 39.Leal MF, Calcagno DQ, Chung J, de Freitas VM, Demachki S, Assumpção PP, et al. Deregulated expression of annexin-A2 and galectin-3 is associated with metastasis in gastric cancer patients. *Clin Exp Med* 2015;15:415-20.
- 40.Emoto K, Sawada H, Yamada Y, Fujimoto H, Takahama Y, Ueno M, et al. Annexin II overexpression is correlated with poor prognosis in human gastric carcinoma. *Anticancer Res* 2001;21:1339-45.
- 41.Sun M-Y, Xing R-H, Gao X-J, Yu X, He H-M, Gao N, et al. ANXA2 regulates the behavior of SGC-7901 cells. *Asian Pac J Cancer Prev* 2013;14:6007-12.
- 42.Zhu F, Xu C, Jiang Z, Jin M, Wang L, Zeng S, et al. Nuclear localization of annexin A1 correlates with advanced disease and peritoneal dissemination in patients with gastric carcinoma. *Anat Rec* 2010;293:1310-4.
- 43.Cheng TY, Wu MS, Lin JT, Lin MT, Shun CT, Huang HY, et al. Annexin A1 is associated with gastric cancer survival and promotes gastric cancer cell invasiveness through the formyl peptide receptor/extracellular signal-regulated kinase/integrin beta₁-binding protein 1 pathway. *Cancer* 2012;118:5757-67.
- 44.Jorge YC, Mataruco MM, Araújo LP, Rossi AFT, De Oliveira JG, Valsechi MC, et al. Expression of annexin-A1 and galectin-1 anti-inflammatory proteins and mRNA in chronic gastritis and gastric cancer. *Mediators Inflammation* 2013;2013.
- 45.Kikuchi S, Kaibe N, Morimoto K, Fukui H, Niwa H, Maeyama Y, et al. Overexpression of Ephrin A2 receptors in cancer stromal cells is a prognostic factor for the relapse of gastric cancer. *Gastric cancer* 2015;18:485-94.
- 46.Zhang ZQ, Li XJ, Liu GT, Xia Y, Zhang XY, Wen H. Identification of Annexin A1 protein expression in human gastric adenocarcinoma using proteomics and tissue microarray. *World J Gastroenterol* 2013;19:7795.
- 47.Gao W, Xu J, Wang F, Zhang L, Peng R, Shu Y, et al. Plasma membrane proteomic analysis of human Gastric Cancer tissues: revealing flotillin 1 as a marker for Gastric Cancer. *BMC Cancer* 2015;15:367.
- 48.Yu S, Li Y, Fan L, Zhao Q, Tan B, Liu Y. Impact of Annexin A3 expression in gastric cancer cells. *Neoplasma* 2014;61:257-64.
- 49.Wang X, Zhang S, Zhang J, Lam E, Liu X, Sun J, et al. Annexin A6 is down-regulated through promoter methylation in gastric cancer. *Am J Transl Res* 2013;5:555.
- 50.Hsu PI, Huang MS, Chen HC, Hsu PN, Lai TC, Wang JL, et al. The significance of ANXA7 expression and its correlation with poor cellular differentiation and enhanced metastatic potential of gastric cancer. *J Surg Oncol* 2008;97:609-14.
- 51.Kim J, Kim MA, Jee CD, Jung EJ, Kim WH. Reduced expression and homozygous deletion of annexin A10 in gastric carcinoma. *Int J Cancer* 2009;125:1842-50.
- 52.Lu SH, Chen YL, Shun CT, Lai JN, Peng SY, Lai PL, et al. Expression and prognostic significance of gastric-specific annexin A10 in diffuse and intestinal-type gastric carcinoma. *J Gastroenterol Hepatol* 2011;26:90-7.
- 53.Thijssen VL, Heusschen R, Caers J, Griffioen AW. Galectin expression in cancer diagnosis and prognosis: A systematic review. *Biochim Biophys Acta* 2015;1855:235-47.
- 54.Bektas S, Bahadır B, Ucan BH, Ozdamar SO. CD24 and galectin-1 expressions in gastric adenocarcinoma and clinicopathologic significance. *Pathol Oncol Res* 2010;16:569-77.
- 55.Chen J, Tang D, Wang S, Li QG, Zhang JR, Li P, et al. High expressions of galectin-1 and VEGF are associated with poor prognosis in gastric cancer patients. *Tumor Biol* 2014;35:2513-9.
- 56.Jung JH, Kim HJ, Yeom J, Yoo C, Shin J, Yoo J, et al. Lowered expression of galectin-2 is associated with lymph node metastasis in gastric cancer. *J Gastroenterol* 2012;47:37-48.
- 57.Ellerhorst J, Troncoso P, Xu XC, Lee J, Lotan R. Galectin-1 and galectin-3 expression in human prostate tissue and prostate cancer. *Urol Res* 1999;27:362-7.
- 58.Van den Brule F, Berchuck A, Bast R, Liu FT, Gillet C, Sobel M, et al. Differential expression of the 67-kD laminin receptor and 31-kD human laminin-binding protein in human ovarian carcinomas. *Eur J Cancer* 1994;30:1096-9.
- 59.Choufani G, Nagy N, Saussez S, Marchant H, Bisschop P, Burchert M, et al. The levels of expression of galectin-1, galectin-3, and the Thomsen-Friedenreich antigen and their

binding sites decrease as clinical aggressiveness increases in head and neck cancers. *Cancer* 1999;86:2353-63.

60. Martínez-Aguilar J, Clifton-Bligh R, Molloy MP. A multiplexed, targeted mass spectrometry assay of the S100 protein family uncovers the isoform-specific expression in thyroid tumours. *BMC Cancer* 2015;15:199.

61. Moss SE, Morgan RO. The annexins. *Genome Biol* 2004;5:219.

62. Liu Y-F, Liu Q-Q, Wang X, Luo C-H. Clinical significance of S100A2 expression in gastric cancer. *Tumor Biology*. 2014;35(4):3731-41.

63. Kang X, Zhang L, Sun J, Ni Z, Ma Y, Chen X, et al. Prohibitin: a potential biomarker for tissue-based detection of gastric cancer. *J Gastroenterol* 2008;43:618-25.

64. Liu T, Tang H, Lang Y, Liu M, Li X. MicroRNA-27a functions as an oncogene in gastric adenocarcinoma by targeting prohibitin. *Cancer Lett* 2009;273:233-42.

65. Leal MF, Cirilo PDR, Mazzotti TKF, Calcagno DQ, Wisniewski F, Demachki S, et al. Prohibitin expression

deregulation in gastric cancer is associated with the 3' untranslated region 1630 C> T polymorphism and copy number variation. *PLoS One* 2014;9:e98583.

66. Balluff B, Rauser S, Meding S, Elsner M, Schöne C, Feuchtinger A, et al. MALDI imaging identifies prognostic seven-protein signature of novel tissue markers in intestinal-type gastric cancer. *Am J Pathol* 2011;179:2720-9.

67. Schwamborn K. Imaging mass spectrometry in biomarker discovery and validation. *Journal of Proteomics*. 2012;75(16):4990-8.

68. Yuan W, Chen Z, Wu S, Ge J, Chang S, Wang X, et al. Expression of EphA2 and E-cadherin in gastric cancer: correlated with tumor progression and lymphogenous metastasis. *Pathology & Oncology Research*. 2009;15(3):473.

69. Ichikawa H, Kanda T, Kosugi Si, Kawachi Y, Sasaki H, Wakai T, et al. Laser microdissection and two-dimensional difference gel electrophoresis reveal the role of a novel macrophage-capping protein in lymph node metastasis in gastric cancer. *J Proteome Res* 2013;12:3780-91.